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### (54) Estrogen receptor

Östrogen-Rezeptor

Récepteur d'oestrogène

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#### Description

**[0001]** This invention relates to the field of receptors belonging to the superfamily of nuclear hormone receptors, in particular to steroid receptors. The invention relates to DNA encoding a novel steroid receptor, the preparation of said receptor, the receptor protein, and the uses thereof.

[0002] Steroid hormone receptors belong to a superfamily of nuclear hormone receptors involved in ligand-dependent transcriptional control of gene expression. In addition, this superfamily consists of receptors for non-steroid hormones such as vitamine D, thyroid hormones and retinoids (Giguère et al, Nature 330, 624-629, 1987; Evans, R.M., Science 240, 889-895,1988). Moreover, a range of nuclear receptor-like sequences have been identified which encode socalled 'orphan' receptors: these receptors are structurally related to and therefore classified as nuclear receptors, although no putative ligands have been identified yet (B.W. O'Malley, Endocrinology 125, 1119-1170, 1989; D.J. Mangelsdorf and R.M. Evans, Cell, 83, 841-850, 1995).

[0003] The superfamily of nuclear hormone receptors share a modular structure in which six distinct structural and functional domains, A to F, are displayed (Evans, Science 240, 889-895, 1988). A nuclear hormone receptor is characterized by a variable N-terminal region (domain A/B), followed by a centrally located, highly conserved DNA-binding domain (hereinafter referred to as DBD; domain C), a variable hinge region (domain D), a conserved ligand-binding domain (herein after referred to as LBD; domain E) and a variable C-terminal region (domain F).

[0004] The N-terminal region, which is highly variable in size and sequence, is poorly conserved among the different members of the superfamily. This part of the receptor is involved in the modulation of transcription activation (Bocquel et al, Nucl. Acid Res., 17, 2581-2595, 1989; Tora et al, Cell 59, 477-487, 1989).

[0005] The DBD consists of approximately 66 to 70 amino acids and is responsible for DNA-binding activity: it targets the receptor to specific DNA sequences called hormone responsive elements (hereinafter referred to as HRE) within the transcription control unit of specific target genes on the chromatin (Martinez and Wahli, In 'Nuclear Hormone Receptors', Acad. Press, 125-153, 1991).

[0006] The LBD is located in the C-terminal part of the receptor and is primarily responsible for ligand binding activity. In this way, the LBD is essential for recognition and binding of the hormone ligand and, in addition possesses a transcription activation function, thereby determining the specificity and selectivity of the hormone response of the receptor. Although moderately conserved in structure, the LBD's are known to vary considerably in homology between the individual members of the nuclear hormone receptor superfamily (Evans, Science 240, 889-895, 1988; P.J. Fuller, FASEB J., 5, 3092-3099, 1991; Mangelsdorf et al, Cell, Vol. 83, 835-839, 1995).

[0007] Functions present in the N-terminal region, LBD and DBD operate independently from each other and it has been shown that these domains can be exchanged between nuclear receptors (Green et al, Nature, Vol. 325, 75-78, 1987). This results in chimeric nuclear receptors, such as described for instance in WO-A-8905355.

[0008] When a hormone ligand for a nuclear receptor enters the cell by diffusion and is recognized by the LBD, it will bind to the specific receptor protein, thereby initiating an allosteric alteration of the receptor protein. As a result of this alteration the ligand/receptor complex switches to a transcriptionally active state and as such is able to bind through the presence of the DBD with high affinity to the corresponding HRE on the chromatin DNA (Martinez and Wahli, 'Nuclear Hormone Receptors',125-153, Acad. Press, 1991). In this way the ligand/receptor complex modulates expression of the specific target genes. The diversity achieved by this family of receptors results from their ability to respond to different ligands.

[0009] The steroid hormone receptors are a distinct class of the nuclear receptor superfamily, characterized in that the ligands are steroid hormones. The receptors for glucocorticoids (GR), mineralcorticoids (MR), progestins (PR), androgens (AR) and estrogens (ER) are classical steroid receptors. Furthermore, the steroid receptors have the unique ability upon activation to bind to palindromic DNA sequences, the so-called HRE's, as homodimers. The GR, MR, PR and AR recognize the same DNA sequence, while the ER recognizes a different DNA sequence. (Beato et al, Cell, Vol. 83, 851-857, 1995). After binding to DNA, the steroid receptor is thought to interact with components of the basal transcriptional machinery and with sequence-specific transcription factors, thus modulating the expression of specific target genes.

[0010] Several HRE's have been identified, which are responsive to the hormone/receptor complex. These HRE's are situated in the transcriptional control units of the various target genes such as mammalian growth hormone genes (responsive to glucocorticoid, estrogen, testosterone), mammalian prolactin genes and progesterone receptor genes (responsive to Estrogen), avian ovalbumin genes (responsive to progesterone), mammalian metallothionein gene (responsive to glucocorticoid) and mammalian hepatic  $\alpha_{2\mu}$ -globulin gene (responsive to estrogen, testosterone, glucocorticoid).

[0011] The steroid hormone receptors have been known to be involved in embryonic development, adult homeostasis as well as organ physiology. Various diseases and abnormalities have been ascribed to a disturbance in the steroid hormone pathway. Since the steroid receptors exercise their influence as hormone-activated transcriptional modulators, it can be anticipated that mutations and defects in these receptors, as well as overstimulation or blocking of these

receptors might be the underlying reason for the attered pattern. A better knowledge of these receptors, their mechanism of action and of the ligands which bind to said receptor might help to create a better insight in the underlying mechanism of the hormone signal transduction pathway, which eventually will lead to better treatment of the diseases and abnormalities linked to altered hormone/receptor functioning.

[0012] For this reason cDNA's of the steroid and several other nuclear receptors of several mammalians, including humans, have been isolated and the corresponding amino acid sequences have been deduced, such as for example the human steroid receptors PR, ER, GR, MR, and AR, the human non-steroid receptors for vitamine D, thyroid hormones, and retinoids such as retinol A and retinoic acid. In addition, cDNA's encoding well over 100 mammalian orphan receptors have been isolated, for which no putative ligands are known yet (Mangelsdorf et al, Cell, Vol.83, 835-839, 1995). However, there is still a great need for the elucidation of other nuclear receptors in order to unravel the various roles these receptors play in normal physiology and pathology.

[0013] The present invention provides for such a novel nuclear receptor. More specifically, the invention provides for an isolated estrogen receptor having an N-terminal domain, a DNA-binding domain, and a ligand-binding domain. wherein the amino acid sequence of said DNA-binding domain exhibits at least 80% homology with the amino acid sequence shown in SEQ ID NO:3 and the amino acid sequence of said ligand-binding domain of said estrogen receptor exhibits at least 70 % homology with the amino acid sequence shown in SEQ ID NO:4,

provided that the estrogen receptor does not have the amino acid sequence:

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MTFYS	PAVMN	YSVPG	STSNL	DGGPV	RLSTS	PNVLW	PTSGH	LSPLA	
THCQS	SLLYA	EPQKS	PWCEA	RSLEH	TLPVN	RETLK	RKLSG	SSCAS	
PVTSP	NAKRD	AHFCP	VCSDY	ASGYH	YGVWS	CEGCK	AFFKR	SIQGH	
NDYIC	PATNQ	CTIDK	NRRKS	CQACR	LRKCY	EVGMV	KCGSR	RERCG	
YRIVR	RQRSS	SEQVH	CLSKA	KRNGG	HAPRV	KELLL	STLSP	EQLVL	
TLLEA	EPPNV	LVSRP	SMPFT	EASMM	MSLTK	LADKE	LVHMI	GWAKK	
IPGFV	ELSLL	DQVRL	LESCW	MEVLM	VGLMW	RSIDH	PGKLI	FAPDL	
VLDRD	EGKCV	EGILE	IFDML	LATTS	RFREL	KLOHK	EYLCV	KAMIL	
LNSSM	YPLAS	ANQEA	ESSRK	LTHLL	NAVTD	ALVWV	IAKSG	ISSQQ	
QSVRL	ANLLM	LLSHV	RHISN	KGMEH	LLSMK	CKNVV	PVYDL	LLEML	
NAHTL	RGYKS	SISGS	ECSST	EDSKN K	ESSO 3	NLOSO.			

The disclaimer relates to the non-prepublished patent application WO 97/09348.

[0014] Hereby, the present invention provides for a novel steroid receptor, having estrogen mediated activity. Said novel steroid receptors are novel estrogen receptors, which are able to bind and be activated by, for example, estradiol, estrone and estriol. According to the present invention it has been found that a novel estrogen receptor is expressed as an 8 kb transcript in human thymus, spleen, peripheral blood lymphocytes (PBLs), ovary and testis. Furthermore, additional transcripts have been identified. Another transcript of approximately 10 kb was identified in ovary, thymus and spleen. In testis, an additional transcript of 1.3 kb was detected. These transcripts are probably generated by alternative splicing of the gene encoding the novel estrogen receptor according to the invention.

[0015] Cloning of the cDNA's encoding the novel estrogen receptors according to the invention revealed that several splicing variants of said receptor can be distinguished. At the protein level, these variants differ only at the C-terminal

[0016] cDNA encoding an ER has been isolated (Green, et al, Nature 320, 134-139, 1986; Greene et al, Science 231, 1150-1154, 1986), and the corresponding amino acid sequence has been deduced. This receptor and the receptor according to the present invention, however, are distinct, and encoded for by different genes with different nucleic acid sequences. Not only do the ER of the prior art (hereinafter referred to as classical ER) and the ER according to the present invention differ in amino acid sequence, they also are located on different chromosomes. The gene encoding the classical ER is located on chromosome 6, whereas the gene encoding the ER according to the invention was found to be located on chromosome 14. The ER according to the invention furthermore distinguishes itself from the classical receptor in differences in tissue distribution, indicating that there may be important differences between these receptors at the level of estrogenic signalling.

[0017] In addition, two orphan receptors, ERRa and ERRS, having an estrogen receptor related structure have been described (Giguère et al, Nature 331, 91-94, 1988). These orphan receptors, however, have not been reported to be able to bind estrodial or any other hormone that binds to the classical ER, and other ligands which bind to these

receptors have not been found yet. The novel estrogen receptor according to the invention distinguishes itself clearly from these receptors since it was found to bind estrogens.

[0018] The fact that a novel ER according to the invention has been found is all the more surprising, since any suggestion towards the existence of additional estrogen receptors was absent in the scientific literature: neither the isolation of the classical ER nor the orphan receptors ERR $\alpha$  and ERR $\beta$  suggested or hinted towards the presence of additional estrogen receptors such as the receptors according to the invention. The identification of additional ER's could be a major step forward for the existing clinical therapies, which are based on the existence of one ER and as such ascribe all estrogen mediated abnormalities and/or diseases to this one receptor. The receptors according to the invention will be useful in the development of hormone analogs that selectively activate either the classical ER or the novel estrogen receptor according to the invention. This should be considered as one of the major advantages of the present invention.

[0019] Thus, in one aspect, the present invention provides for isolated cDNA encoding a novel steroid receptor. In particular, the present invention provides for isolated cDNA encoding the novel estrogen receptor as defined above.

[0020] According to this aspect of the present invention, there is provided an isolated DNA encoding a steroid receptor protein having an N-terminal domain, a DNA-binding domain and a ligand-binding domain, wherein the amino acid sequence of said DNA-binding domain of said receptor protein exhibits at least 80% homology with the amino acid sequence shown in SEQ ID NO:3, and the amino acid sequence of said ligand-binding domain of said receptor protein exhibits at least 70% homology with the amino acid sequence shown in SEQ ID NO:4.

[0021] In particular, the isolated DNA encodes a steroid receptor protein having an N-terminal domain, a DNA-binding domain and a ligand-binding domain, wherein the amino acid sequence of said DNA-binding domain of said receptor protein exhibits at least 90%, preferably 95%, more preferably 98%, most preferably 100% homology with the amino acid sequence shown in SEQ ID NO:3.

[0022] More particularly, the isolated DNA encodes a steroid receptor protein having an N-terminal domain, a DNA-binding domain and a ligand-binding domain, wherein the amino acid sequence of said ligand-binding domain of said receptor protein exhibits at least 75%, preferably 80%, more preferably 90%, most preferably 100% homology with the amino acid sequence shown in SEQ ID NO:4.

[0023] A preferred isolated DNA according to the invention encodes a steroid receptor protein having the amino acid sequence shown in SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:21 or SEQ ID NO:25.

[0024] A more preferred isolated DNA according to the invention is an isolated DNA comprising a nucleotide sequence shown in SEQ ID NO:1, SEQ ID NO:2, SEQ ID NO:20 or SEQ ID NO:24.

[0025] The DNA according to the invention may be obtained from cDNA. Alternatively, the coding sequence might be genomic DNA, or prepared using DNA synthesis techniques.

[0026] The DNA according to the invention will be very useful for in vivo expression of the novel receptor proteins according to the invention in sufficient quantities and in substantially pure form.

[0027] In another aspect of the invention, there is provided for a steroid receptor comprising the amino acid sequence encoded by the above described DNA molecules.

[0028] The steroid receptor according to the invention has an N-terminal domain, a DNA-binding domain and a ligand-binding domain, wherein the amino acid sequence of said DNA-binding domain of said receptor exhibits at least 80% homology with the amino acid sequence shown in SEQ ID NO:3, and the amino acid sequence of said ligand-binding domain of said receptor exhibits at least 70% homology with the amino acid sequence shown in SEQ ID NO:4.

[0029] In particular, the steroid receptor according to the invention has an N-terminal domain, a DNA-binding domain and a ligand-binding domain, wherein the amino acid sequence of said DNA-binding domain of said receptor exhibits at least 90%, preferably 95%, more preferably 98%, most preferably 100% homology with the amino acid sequence shown in SEQ ID NO:3.

[0030] More particular, the steroid receptor according to the invention has an N-terminal domain, a DNA-binding domain and a ligand-binding domain, wherein the amino acid sequence of said ligand-binding domain of said receptor exhibits at least 75%, prefearbly 80%, more preferably 90%, most preferably 100% homology with the amino acid sequence shown in SEQ ID NO:4.

[0031] It will be clear for those skilled in the art that also steroid receptor proteins comprising combined DBD and LBD preferences and DNA encoding such receptors are subject of the invention.

[0032] Preferably, the steroid receptor according to the Invention comprises an amino acid sequence shown in SEQ ID NO:5, SEQ ID NO:6, SEQ ID NO:21 or SEQ ID NO:25.

[0033] Also within the scope of the present invention are steroid receptor proteins which comprise variations in the amino acid sequence of the DBD and LBD without loosing their respective DNA-binding or ligand-binding activities.

The variations that can occur in those amino acid sequences comprise deletions, substitutions, insertions, inversions or additions of (an) amino acid(s) in said sequence, said variations resulting in amino acid difference(s) in the overall sequence. It is well known in the art of proteins and peptides that these amino acid differences lead to amino acid sequences that are different from, but still homologous with the native amino acid sequence they have been derived

from.

[0034] Amino acid substitutions that are expected not to essentially alter biological and immunological activities, have been described in for example Dayhof, M.D., Atlas of protein sequence and structure, Nat. Biomed. Res. Found., Washington D.C., 1978, vol. 5, suppl. 3. Amino acid replacements between related amino acids or replacements which have occurred frequently in evolution are, inter alia Ser/Ala, Ser/Gly, Asp/Gly, Arg/Lys, Asp/Asn, Ile/Val. Based on this information Lipman and Pearson developed a method for rapid and sensitive protein comparison (Science 227, 1435-1441, 1985) and determining the functional similarity between homologous polypeptides.

[0035] Variations in amino acid sequence of the DBD according to the invention resulting in an amino acid sequence that has at least 80% homology with the sequence of SEQ ID NO:3 will lead to receptors still having sufficient DNA binding activity. Variations in amino acid sequence of the LBD according to the invention resulting in an amino acid sequence that has at least 70% homology with the sequence of SEQ ID NO:4 will lead to receptors still having sufficient ligand binding activity.

[0036] Homology as defined herein is expressed in percentages, determined via PCGENE. Homology is calculated as the percentage of identical residues in an alignment with the sequence according to the invention. Gaps are allowed to obtain maximum alignment.

[0037] Comparing the amino acid sequences of the classical ER and the ER's according to the invention revealed a high degree of similarity within their respective DBD's. The conservation of the P-box (amino acids E-G-X-X-A) which is responsible for the actual interactions of the classical ER with the target DNA element (Zilliacus et al., Mol.Endo. 9, 389, 1995; Glass, End.Rev. 15, 391, 1994), is indicative for a recognition of estrogen responsive elements (ERE's) by the ER's according to the invention. The receptors according to the invention indeed showed ligand-dependent transactivation on ERE-containing reporter constructs. Therefore, the classical ER and the novel ER's according to the invention may have overlapping target gene specificities. This could indicate that in tissues which co-express both respective ER's, these receptors compete for ERE's. The ER's according to the invention may regulate transcription of target genes differently from classical ER regulation or could simply block classical ER functioning by occupying estrogen responsive elements. Alternatively, transcription might be influenced by heterodimerization of the different receptors.

[0038] Thus, a preferred steroid receptor according to the invention comprises the amino acid sequence E-G-X-X-A within the P box of the DNA binding domain, wherein X stands for any amino acid. Also within the scope of the invention is isolated DNA encoding such a receptor.

[0039] Methods to prepare the receptors according to the invention are well known in the art (Sambrook et al., Molecular Cloning: a Laboratory Manual, Cold Spring Harbor Laboratory Press, Cold Spring Harbor, 1989). The most practical approach is to produce these receptors by expression of the DNA encoding the desired protein.

[0040] A wide variety of host cell and cloning vehicle combinations may be usefully employed in cloning the nucleic acid sequence coding for the receptor of the invention. For example, useful cloning vehicles may include chromosomal, non-chromosomal and synthetic DNA sequences such as various known bacterial plasmids and wider host range plasmids and vectors derived from combinations of plasmids and phage or virus DNA. Useful hosts may include bacterial hosts, yeasts and other fungi, plant or animal hosts, such as Chinese Hamster Ovary (CHO) cells or monkey cells and other hosts (with exception of linman beings).

[0041] Vehicles for use in expression of the ligand-binding domain of the present invention will further comprise control sequences operably linked to the nucleic acid sequence coding for the ligand-binding domain. Such control sequences generally comprise a promoter sequence and sequences which regulate and/or enhance expression levels. Furthermore an origin of replication and/or a dominant selection marker are often present in such vehicles. Of course control and other sequences can vary depending on the host cell selected.

[0042] Techniques for transforming or transfecting host cells are quite known in the art (see, for instance, Sambrook et al., Molecular Cloning: A Laboratory Manual, Cold Spring Harbor Laboratory, 1989).

[0043] Recombinant expression vectors comprising the DNA of the invention as well as cells transformed with said DNA or said expression vector also form part of the present invention.

**[0044]** Since steroid receptors have three domains with different functions, which are more or less independent, it is possible that all three functional domains have been derived from different members of the steroid receptor superfamily.

[0045] Molecules which contain parts having a different origin are called chimeric. Such a chimeric receptor comprising the ligand-binding domain and/or the DNA-binding domain of the invention may be produced by chemical linkage, but most preferably the coupling is accomplished at the DNA level with standard molecular biological methods by fusing the nucleic acid sequences encoding the necessary steroid receptor domains. Hence, DNA encoding the chimeric receptor proteins according to the invention are also subject of the present invention.

[0046] Such chimeric proteins can be prepared by transfecting DNA encoding these chimeric receptor proteins to suitable host cells and culturing these cells under suitable conditions.

[0047] It is extremely practical if, next to the information for the expression of the steroid receptor, also the host cell

is transformed or transfected with a vector which carries the information for a reporter molecule. Such a vector coding for a reporter molecule is characterized by having a promoter sequence containing one or more hormone responsive elements (HRE) functionally linked to an operative reporter gene. Such a HRE is the DNA target of the activated steroid receptor and, as a consequence, it enhances the transcription of the DNA coding for the reporter molecule. In *in vivo* settings of steroid receptors the reporter molecule comprises the cellular response to the stimulation of the ligand. However, it is possible *in vitro* to combine the ligand-binding domain of a receptor to the DNA binding domain and transcription activating domain of other steroid receptors, thereby enabling the use of other HRE and reporter molecule systems. One such a system is established by a HRE presented in the MMTV-LTR (mouse mammary tumor virus long terminal repeat sequence in connection with a reporter molecule like the firefly luciferase gene or the bacterial gene for CAT (chloramphenicol transferase). Other HRE's which can be used are the rat oxytocin promotor, the retinoic acid responsive element, the thyroid hormone responsive element, the estrogen responsive element and also synthetic responsive elements have been described (for instance in Fuller, ibid. page 3096). As reporter molecules next to CAT and luciferase β-galactosidase can be used.

[0048] Steroid hormone receptors and chimeric receptors according to the present invention can be used for the *in vitro* identification of novel ligands or hormonal analogs. For this purpose binding studies can be performed with cells transformed with DNA according to the invention or an expression vector comprising DNA according to the invention, said cells expressing the steroid receptors or chimeric receptors according to the invention.

**[0049]** The novel steroid hormone receptor and chimeric receptors according to the invention as well as the ligand-binding domain of the invention, can be used in an assay for the identification of functional ligands or hormone analogs for the nuclear receptors.

[0050] Thus, the present invention provides for a method for identifying functional ligands for the steroid receptors and chimeric receptors according to the invention, said method comprising the steps of

- a) introducing into a suitable host cell 1) DNA or an expression vector according to the invention, and 2) a suitable reporter gene functionally linked to an operative hormone response element, said HRE being able to be activated by the DNA-binding domain of the receptor protein encoded by said DNA;
- b) bringing the host cell from step a) into contact with potential ligands which will possibly bind to the ligand-binding domain of the receptor protein encoded by said DNA from step a);
- c) monitoring the expression of the receptor protein encoded by said reporter gene of step a).

[0051] If expression of the reporter gene is induced with respect to basic expression (without ligand), the functional ligand can be considered as an agonist; if expression of the reporter gene remains unchanged or is reduced with respect to basic expression, the functional ligand can be a suitable (partial) antagonist.

[0052] For performing such kind of investigations host cells which have been transformed or transfected with both a vector encoding a functional steroid receptor and a vector having the information for a hormone responsive element and a connected reporter molecule are cultured in a suitable medium. After addition of a suitable ligand, which will activate the receptor the production of the reporter molecule will be enhanced, which production simply can be determined by assays having a sensitivity for the reporter molecule. See for instance WO-A-8803168. Assays with known steroid receptors have been described (for instance S. Tsai et al., Cell 57, 443, 1989; M. Meyer et al., Cell 57, 433, 1989).

#### Legends to the figures

#### Figure 1.

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[0053] Northern analysis of the novel estrogen receptor (ER $\beta$ ). Two different multiple tissue Northern blots (Clontech) were hybridised with a specific probe for ER $\beta$  (see examples). Indicated are the human tissues the RNA originated from and the position of the size markers in kilobases (kb).

### Figure 2.

[0054] Histogram showing the 3- to 4-fold stimulatory effect of 17β-estradiol, estriol and estrone on the luciferase activity mediated by ERβ. An expression vector encoding ERβ was transiently transfected into CHO cells together with a reporter construct containing the rat oxytocin promoter in front of the firefly luciferase encoding sequence (see examples).

#### Figure 3.

[0055] Effect of 17β-estradiol (E2) alone or in combination with the anti-estrogen ICI-164384 (ICI) on ERα and ERβ.

Expression constructs for ER $\alpha$  (the classical ER) and ER $\beta$  were transiently transfected into CHO cells together with the rat oxytocin promoter-luciferase reporter construct described in the examples. Luciferase activities were determined in triplicate and normalised for transfection efficiency by measuring  $\beta$ -galactosidase in the same lysate.

#### Figure 4.

[0056] Expression of ER $\alpha$  and ER $\beta$  in a number of cell lines determined by RT-PCR analysis (see examples). The cell lines used were derived from different tissues/cell types: endometrium (ECC1, Ishikawa, HEC-1A, RL95-2); osteosarcoma (SAOS-2, U2-OS, HOS, MG63); breast tumours (MCF-7, T47D), endothelium (HUV-EC-C, BAEC-1); smooth muscle (HISM, PAC-1, A7R5, A10, RASMC, CavaSMC); liver (HepG2); colon (CaCo2); and vagina (Hs-760T, SW-954). [0057] All cell lines were human except for PAC-1, A7R5, A10 and RASMC which are of rat origin, BAEC-1 which is of bovine origin and CavaSMC which is of guinea pig origin.

#### Figure 5.

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[0058] Transactivation assay using stably transfected CHO cell lines expressing ER $\alpha$  or ER $\beta$  together with the rat oxytocin-luciferase estrogen-responsive reporter (see examples for details). Hormone-dependent transactivation curves were determined for 17 $\beta$ -estradiol and for Org4094. For the ER antagonist raloxifen, cells were treated with 2 x 10<sup>-10</sup> mol/L 17 $\beta$ -estradiol together with increasing concentrations of raloxifen. Maximal values of the responses were arbitrarily set at 100%.

#### Examples

### A. Molecular cloning of the novel estrogen receptor.

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[0059] Two degenerate oligonucleotides containing inosines (I) were based on conserved regions of the DNA-binding domains and the ligand-binding domains of the human steroid hormone receptors.

[0060] Primer #1:

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5'-GGIGA (C/T) GA (A/G) GC (A/T) TCIGGITG (C/T) CA (C/T) TA (C/T) GG-3'

(SEQ ID NO:7).

[0061] Primer #2:

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### 5'-AAGCCTGG(C/G)A(C/T)IC(G/T)(C/T)TTIGCCCAI(C/T)TIAT-3'

(SEQ ID NO:8).

[0062] As template, cDNA from human EBV-stimulated PBLs (peripheral blood leukocytes) was used. One microgram of total RNA was reverse transcribed in a 20 µl reaction containing 50 mM KCl, 10 mM Tris-HCl pH 8.3, 4 mM MgCl2, 1 mM dNTPs (Pharmacia), 100 pmol random hexanucleotides (Pharmacia), 30 Units RNAse inhibitor (Pharmacia) and 200 Units M-MLV Reverse transcriptase (Gibco BRL). Reaction mixtures were incubated at 37°C for 30 minutes and heat-inactivated at 100°C for 5 minutes. The cDNA obtained was used in a 100 µl PCR reaction containing 10 mM Tris-HCl pH 8.3, 50 mM KCl, 1.5 mM MgCl2, 0.001% gelatin (w/v), 3% DMSO, 1 microgram of primer #1 and primer #2 and 2.5 Units of Amplitaq DNA polymerase (Perkin Elmer). PCR reactions were performed in the Perkin Elmer 9600 thermal cycler. The initial denaturation (4 minutes at 94°C) was followed by 35 cycles with the following conditions: 30 sec. 94°C, 30 sec. 45°C, 1 minute 72°C and after 7 minutes at 72°C the reactions were stored at 4°C. Aliquots of these reactions were analysed on a 1.5% agarose gel. Fragments of interest were cut out of the gel, reamplified using identical PCR-conditions and purified using Qiaex II (Qiagen). Fragments were cloned in the pCRII vector and transformed into bacteria using the TA-cloning kit (Invitrogen). Plasmid DNA was isolated for nucleotide sequence analysis using the Qiagen plasmid midi protocol (Qiagen). Nucleotide sequence analysis was performed with the ALF automatic sequencer (Pharmacia) using a T7 DNA sequencing kit (Pharmacia) with vector-specific or fragment-specific primers.

[0063] One cloned fragment corresponded to a novel estrogen receptor (ER) which is closely related to the classical estrogen receptor. Part of the cloned novel estrogen receptor fragment (nucleotides 466 to 797 in SEQ ID 1) was amplified by PCR using oligonucleotide #3 TGTTACGAAGTGGGAATGGTGA (SEQ ID NO:9) and oligonucleotide #2 and used as a probe to screen a human testis cDNA library in λgt11 (Clontech #HL1010b). Recombinant phages were

plated (using Y1090 bacteria grown in LB medium supplemented with 0.2% maltose) at a density of 40.000 pfu (plaqueforming units) per 135 mm dish and replica filters (Hybond-N, Amersham) were made as described by the supplier. Filters were prehybridised in a solution containing 0.5 M phosphate buffer (pH 7.5) and 7% SDS at 65°C for at least 30 minutes. DNA probes were purified with Qiaex II (Qiagen), 32P-labeled with a Decaprime kit (Ambion) and added to the prehybridisation solution. Filters were hybridised at 65°C overnight and then washed in 0.5 X SSC/0.1% SDS at 65°C. Two positive plaques were identified and could be shown to be identical. These clones were purified by rescreening one more time. A PCR reaction on the phage eluates with the λgt11-specific primers #4: 5'-TTGACACCA-GACCAACTGGTAATG-3' (SEQ ID NO:10) and #5: 5'-GGTGGCGACGACTCCTGGAGCCCG-3' (SEQ ID NO:11) yielded a fragment of 1700 basepairs on both clones. Subsequent PCR reactions using combinations of a gene-specific primer #6: 5'-GTACACTGATTTGTAGCTGGAC-3' (SEQ ID NO:12) with the λgt11 primer #4 and gene-specific primer #7; 5'-CCATGATGATGTCCCTGACC-3' (SEQ ID NO:13) with λgt11 primer primer #5 yielded fragments of approximately 450 bp and 1000 bp, respectively, which were cloned in the pCRII vector and used for nucleotide sequence analysis. The conditions for these PCR reactions were as described above except for the primer concentrations (200 ng of each primer) and the annealing temperature (60°C). Since in the cDNA clone the homology with the ER is lost abruptly at a site which corresponds to the exon 7/exon 8 boundary in the ER (between nucleotides 1247 and 1248 in SEQ ID NO:1), it was suggested that this sequence corresponds to intron 7 of the novel ER gene. For verification of the nucleotide sequences of this cDNA clone, a 1200 bp fragment was generated on the cDNA clone with \(\lambda\gamma\)11 primer #4 with a gene-specific primer #8 corresponding to the 3' end of exon 7: 5'-TCGCATGCCTGACGTGGGAC-3' (SEQ ID NO:14) using the proofreading Pfu polymerase (Stratagene). This fragment was also cloned in the pCRII vector and completely sequenced and was shown to be identical to the sequences obtained earlier.

[0064] To obtain nucleotide sequences of the novel ER downstream of exon 7, a degenerate oligonucleotide based on the AF-2 region of the classical ER (#9: 5'-GGC(C/G)TCCAGCATCTCCAG(C/G)A(A/G)CAG-3'; SEQ ID NO:15) was used together with the gene-specific oligonucleotide #10: 5'-GGAAGCTGGCTCACTTGCTG-3' (SEQ ID NO:16) using testis cDNA as template (Marathon ready testis cDNA, Clontech Cat #7414-1). A specific 220 bp fragment corresponding to nucleotides 1112 to 1332 in SEQ ID No. 1 was cloned and sequenced. Nucleotides 1112 to 1247 were identical to the corresponding sequence of the cDNA clone. The sequence downstream thereof is highly homologous with the corresponding region in the classical ER. In order to obtain sequences of the novel ER downstream of the AF-2 region, RACE (rapid amplification of cDNA ends) PCR reactions were performed using the Marathon-ready testis cDNA (Clontech) as template. The initial PCR was performed using oligonucleotide #11: 5'-TCTTGTTCTGGACAG-GGATG-3' (SEQ ID NO:17) in combination with the AP1 primer provided in the kit. A nested PCR was performed on an aliquot of this reaction using oligonucleotide #10 (SEQ ID NO:16) in combination with the oligo dT primer provided in the kit. Subsequently, an aliquot of this reaction was used in a nested PCR using oligonucleotide #12:5'-GCATGGAA-CATCTGCTCAAC-3' (SEQ ID NO:18) in combination with the oligo dT primer. Nucleotide sequence analysis of a specific fragment that was obtained (corresponding to nucleotides 1256 to 1431 in SEQ ID NO 1) revealed a sequence encoding the carboxyterminus of the novel ER ligand-binding domain, including an F-domain and a translational stop codon and part of the 3' untranslated sequence which is not included in SEQ ID NO:1. The deduced amino acid sequence is shown in SEQ ID NO:5.

[0065] In order to investigate the possibility that the novel estrogen receptor had additional, upstream translation-initiation codons, RACE-PCR experiments were performed using Marathon-ready testis cDNA (Clontech Cat. # 7414-1). First a PCR was performed using oligonucleotide SEQ ID NO:12 (antisense corresponding to nucleotides 416-395 in SEQ ID NO:1) and AP-1 (provided in the kit). A nested PCR was then performed using oligonucleotide having SEQ ID NO:27 (antisense corresponding to nucleotides 254-231 in SEQ ID NO:1) with AP-2 (provided in the kit). From the smear that was obtained, the region corresponding to fragments larger than 300 basepairs was cut out, purified using the GenecleanII kit (Bio101) and cloned using the TA-cloning kit (Clontech). Colonies were screened by PCR using gene-specific primers: SEQ ID NO:22 and SEQ ID NO:28. The clone containing the largest insert was sequenced. The nucleotide sequence corresponds to nucleotides 1 to 490 in SEQ ID NO:24. It is clear from this sequence that the first in-frame upstream translation initiation codon is present at position 77-79 in SEQ ID NO:24. Upstream of this translational startcodon an in-frame stop-codon is present (11-13 in SEQ ID NO:24). Consequently, the reading frame of the novel estrogen receptor is 530 amino acids (shown in SEQ ID NO:25) and has a calculated molecular mass of 59.234 kD.

[0066] To confirm the nucleotide sequences obtained by 5' RACE, human genomic clones were obtained and analysed. A human genomic library in λEMBL3 (Clontech HL1067J) was screened with a probe corresponding to nucleotides 1 to 416 in SEQ ID NO:1. A strongly hybridizing clone was plaque-purified and DNA was isolated using standard protocols (Sambrook et al, 1989). The DNA was digested with several restriction enzymes, electrophoresed on agarose gel and blotted onto Nylon filters. Hybridisation of the blot with a probe corresponding to the above-mentioned RACE fragment (nucleotides 1-490 in SEQ ID NO:24) revealed a hybridizing Sau3A fragment of approximately 800 basepairs. This fragment was cloned into the BamH1 site of pGEM3Z and sequenced. The nucleotide sequence contained one base difference which is probably a PCR-induced point mutation in the RACE fragment. Nucleotide 172 was a G residue

in the 5'RACE fragment, but an A residue in several independent genomic subclones.

#### B. Identification of two splice variants of the novel estrogen receptor.

[0067] Rescreening of the testis cDNA library with a probe corresponding to nucleotides 918 to 1246 in SEQ ID No. 1 yielded two hybridizing clones, the 3' end of which were amplified by PCR (gene-specific primer #10: 5'-GGAAGCT-GGCTCACTTGCTG-3' (SEQ ID NO:16) together with primer #4, SEQ ID NO:10), cloned and sequenced. One clone was shown to contain an alternative exon 8 (exon 8B) of the novel ER. In SEQ ID No. 2 the protein encoding part and the stopcodon of this splice variant are presented. As a consequence of the introduction of this exon through an alternative splicing reaction, the reading frame encoding the novel ER is immediately terminated, thereby creating a truncation of the carboxyterminus of the novel ER (SEQ ID NO:6).

[0068] Screening of a human thymus cDNA library (Clontech HL1074a) with the probe corresponding to nucleotides 918 to 1246 in SEQ ID No. 1, revealed another splice variant. The 3' end of one hybridizing clone was amplified using primer #10 (SEQ-ID NO:16) with the \(\lambda\)gt10-specific primer #13 5'-AGCAAGTTCAGCCTGTTAAGT-3' (SEQ ID NO:19), cloned and sequenced. The obtained nucleotide sequence upstream of the exon 7/exon 8 boundary was identical to the clones identified earlier. However, an alternative exon 8 (exon 8C) was present at the 3' end encoding two C-terminal amino acids followed by a stop-codon. The nucleotide sequence of the protein-encoding part of this splice variant is shown in SEQ ID NO:20, the corresponding protein sequence is SEQ ID NO:21.

[0069] These two variants of the novel estrogen receptor do not contain the AF-2 region and therefore probably lack the ability to modulate transcription of target genes in a ligand-dependent fashion. However, the variants potentially could interfere with the functioning of the wild-type classical ER and/or the wild-type novel ER, either by heterodimerization or by occupying estrogen response elements or by interactions with other transcription factors. A mutant of the classical ER (ER1-530) has been described which closely resembles the two variants of the novel estrogen receptor described above. ER1-530 has been shown to behave as a dominant-negative receptor i.e. it can modulate the intracellular activity of the wild type ER (Ince et al, J. Biol. Chem. 268, 14026-14032, 1993).

#### C. Northern blot analysis.

[0070] Human multiple tissue Northern blots (MTN-blots) were purchased from Clontech and prehybridized for at least 1 hour at 65°C in 0.5 M phosphate buffer pH 7.5 with 7% SDS. The DNA fragment that was used as a probe (corresponding to nucleotides 466 to 797 in SEQ ID No. 1) was <sup>32</sup>P-labeled using a labelling kit (Ambion), denatured by boiling and added to the prehybridisation solution. Washing conditions were: 3X SSC at room temperature, followed by 3 X SSC at 65°C, and finally 1 X SSC at 65°C. The filters were than exposed to X-ray films for one week. Two transcripts of approximately 8 kb and 10 kb were detected in thymus, spleen, ovary and testis. In addition, a 1.3 kb transcript was detected in testis.

### D. RT-PCR analysis of expression of ERa and ERB in cell lines.

[0071] RNA was isolated from a number of human and animal cell lines using RNAzol B (Cinna/Biotecx). cDNA was made using 2.5 microgram of total RNA using the Superscript II kit (BRL) following the manufacturers instructions. A portion of the cDNA was used for specific PCR amplifications of fragments corresponding either to mRNA encoding the ER or to the novel estrogen receptor. (It should be emphasized that the primers used are based on human and rat sequences, whereas some of the cell lines were not rat or human, see legend of Figure 4). Primers used were for ERα: sense 5'-GATGGGCTTACTGACCAACC-3' and antisense 5'-AGATGCTCCATGCCTTTG-3' generating a 548 base pair fragment corresponding to part of the LBD. For ERβ: sense 5'-TTCACCGAGGCCTCCATGATG-3' and antisense 5'-CA-GATGTTCCATGCCCTTGTT-3' generating a 565 base pair fragment corresponding to part of the LBD. The PCR samples were analysed on agarose which were blotted onto Nylon membranes. These blots were hybridised with <sup>32</sup>P-labeled PCR fragments, generated with the above-mentioned primers on ERα and ERβ plasmid DNA using standard experimental procedures (Sambrook *et al*, 1989).

### E. Ligand-dependent transcription activation by the novel estrogen receptor protein.

### Cell culture

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[0072] Chinese Hamster Ovary (CHO K1) cells were obtained from ATCC (CCL61) and maintained at 37°C in a humidified atmosphere (5% CO<sub>2</sub>) as a monolayer culture in fenoired-free M505 medium. The latter medium consists of a mixture (1:1) of Dulbecco's Modified Eagle's Medium (DMEM, Gibco 074-200) and Nutrient Medium F12 (Ham's F12, Gibco 074-1700) supplemented with 2.5 mg/ml sodium carbonate (Baker), 55 µg/ml sodium pyruvate (Fluka), 2.3

μg/ml β-mercaptoethanol (Baker), 1.2 μg/ml ethanolamine (Baker), 360 μg/ml L-glutamine (Merck), 0.45 μg/ml sodium selenite (Fluka), 62.5 μg/ml penicillin (Mycopharm), 62.5 μg/ml streptomycin (Serva), and 5% charcoal-treated bovine calf serum (Hyclone).

#### 5 Recombinant vectors

[0073] The ERβ-encoding sequence as presented in SEQ ID No. 1 was amplified by PCR using oligonucleotides 5'-CTTGGATCCATAGCCCTGCTGATGAATTACAG-3' (SEQ ID NO:22 underlined is the translation initiation codon) in combination with 5'-GATGGATCCTCACCTCAGGGCCAGGCGTCACTG-3' (SEQ ID NO:23) (underlined is the translation stopcodon, antisense). The resulting BamH1 fragment (approximately 1450 base pairs) were then cloned in the mammalian cell expression vector pNGV1 (Genbank accession No. X99274).

[0074] An expression construct encoding the ERβ reading frame as presented in SEQ ID NO:24 was made by replacing a BamH1-Msc1 fragment (nucleotides 1-81 in SEQ ID No. 1) by a BamH1-Msc1 fragment corresponding to nucleotides 77-316 in SEQ ID No. 24. The latter fragment was made by PCR with SEQ ID NO:26 in combination with SEQ ID NO:28 using the above mentioned 5' RACE fragment.

[0075] The reporter vector was based on the rat oxytocin gene regulatory region (position -363/+16 as a HindIII/ Mbol fragment; R.Iveli, and D.Richter, Proc.Natl.Acad.Sci.USA <u>81</u>, 2006-2010, 1984) linked to the firefly luciferase encoding sequence; the regulatory region of the oxytocin gene was shown to possess functional estrogen hormone response elements *in vitro* for both the rat (R.Adan *et al*, Biochem.Biophys.Res.Comm. <u>175</u>, 117-122, 1991) and the human (S.Richard, and H.Zingq, J.Biol.Chem. 265, 6098-6103, 1990).

### Transient transfection

[0076] 1 x 10 $^{5}$  CHO cells were seeded in 6-wells Nunclon tissue culture plates and DNA was introduced by use of lipofectin (Gibco BRL). Hereto, the DNA (1  $\mu$ g of both receptor and reporter vector in 250  $\mu$ L Optimem, Gibco BRL) was mixed with an equal volume of lipofectin reagent (7  $\mu$ L in 250  $\mu$ L Optimem, Gibco) and allowed to stand at room temperature for 15 min. After washing the cells twice with serum-free medium (M505) new medium (500  $\mu$ L Optimem, Gibco) was added to the cells followed by the dropwise addition of the DNA-lipofectin mixture. After incubation for a 5 hour period at 37 $^{\circ}$ C cells were washed twice with fenolred-free M505 + 5% charcoal-treated bovine calf serum and incubated overnight at 37 $^{\circ}$ C. After 24 hours hormones were added to the medium (10 $^{-7}$  mol/L). Cell extracts were made 48 hours posttransfection by the addition of 200  $\mu$ L lysisbuffer (0.1 M phosphate buffer pH7.8, 0.2% Triton X-100). After incubation for 5 min at 37 $^{\circ}$ C the cell suspension was centrifuged (Eppendorf centrifuge, 5 min) and 20  $\mu$ L sample was added to 50  $\mu$ L luciferase assay reagent (Promega). Light emission was measured in a luminometer (Berthold Biolumat) for 10 sec at 562 nm.

### Stable transfection of the novel estrogen receptor.

[0077] The expression plasmid encoding full-length ERβ1-530 (see above) was stably transfected in CHO K1 cells as previously described (Theunissen *et al.*, J. Biol. Chem. 268, 9035-9040, 1993). Single cell clones that were obtained this way were screened by transient transfection of the reporter plasmid (rat oxytocin-luciferase) as described above. Selected clones were used for a second stable transfection of the rat oxytocin-luciferase reporter plasmid together with the plasmid pDR2A which contains a hygromycine resitance gene for selection. Single cell clones obtained were tested for a response to 17β-estradiol. Subsequently, a selected single cell clone was used for transactivation studies. Briefly, cells were seeded in 96-wells at (1.6x10<sup>4</sup> cells per well). After 24 hours different concentrations of hormone were diluted in medium and added to the wells. For antagonistic experiments, 2x10<sup>-10</sup> M. 17β-estradiol was added to each well and different concentrations of antagonists were added. Cells were washed once with PBS after a 24 hour incubation and then lysed by the addition of 40 microliter lysis buffer (see above). Luciferase reagent was added (50 microliter) to each well and light emission was measured using the Topcount (Packard).

### 50 Results.

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[0078] A comparison of the two expression constructs (SEQ ID NO:1 and SEQ ID NO:24) in transient transfections in CHO cells showed identical transactivation in response to a number of agonists and antagonists. CHO cells transiently transfected with ER $\beta$  expression vector and a reporter plasmid showed a 3 to 4 fold increase in luciferase activity in response to 17 $\beta$ -estradiol as compared to untreated cells (see Figure 2). A similar transactivation was obtained upon treatment with estriol and estrone. The results indicate not only that the novel ER (ER $\beta$ ) can bind estrogen hormones but also that the ligand-activated receptor can bind to the estrogen-response elements (EREs) within the rat oxytocin promoter and activate transcription of the luciferase reporter gene. Figure 3 shows that in an independent similar

experiment  $10^{-9}$  mol/L  $17\beta$ -estradiol gave an 18-fold stimulation with ER $\alpha$  and a 7-fold stimulation with ER $\beta$ . In addition, the antiestrogen ICI-164384 was shown to be an antagonist for both ER $\alpha$  and ER $\beta$  when activated with  $17\beta$ -estradiol, whereas the antagonist alone had no effect. In this experiment 0.25  $\mu$ g  $\beta$ -galactosidase vector was co-transfected in order to normalize for differences in transfection efficiency.

[0079] Transactivation studies performed on stably transfected ER $\alpha$  and ER $\beta$  cell lines gave similar absolute luciferase values. The curves for 17 $\beta$ -estradiol are very similar and show that half-maximal transactivation is reached with lower concentrations of hormone on ER $\alpha$  as compared to ER $\beta$  (Figure 5). For Org4094 this is also the case however, the effect observed is much more pronounced. The curves for raioxifen show that the potency of this antagonist to block transactivation on ER $\alpha$  is greater compared to its potency to block ER $\beta$  transactivation.

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SEQUENCE LISTING

#### [0080]

- (1) GENERAL INFORMATION:
  - (i) APPLICANT:
    - (A) NAME: Akzo nobel n.v.
    - (B) STREET: Velperweg 76
    - (C) CITY: Arnhem
    - (E) COUNTRY: The Netherlands
    - (F) POSTAL CODE (ZIP): 6824 BM
    - (G) TELEPHONE: 0412-666379
    - (H) TELEFAX: 0412-650592
    - (I) TELEX: 37503 akpha nl
  - (ii) TITLE OF INVENTION: Novel estrogen receptor
- 30 (iii) NUMBER OF SEQUENCES: 28
  - (iv) COMPUTER READABLE FORM:
    - (A) MEDIUM TYPE: Floppy disk
    - (B) COMPUTER: IBM PC compatible
    - (C) OPERATING SYSTEM: PC-DOS/MS-DOS
    - (D) SOFTWARE: Patentin Release #1.0, Version #1.30 (EPO)
  - (2) INFORMATION FOR SEQ ID NO: 1:
- 40

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- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 1434 base pairs
  - (B) TYPE: nucleic acid
  - (C) STRANDEDNESS: double
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: cDNA
- 50 (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 1:

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10	CAGTTATCAC	ATCTGTATGC	GGAACCTCAA	AAGAGTCCCT	GGTĢTGAAGO	AAGATCGCTA	180
,0	GAACACACCT	TACCTGTAAA	CAGAGAGACA	CTGAAAAGGA	AGGTTAGTGG	GAACCGTTGC	240
15	GCCAGCCCTG	TTACTGGTCC	AGGTTCAAAG	AGGGATGCTC	ACTTCTGCGC	TGTCTGCAGC	300
	GATTACGCAT	CGGGATATCA	CTATGGAGTC	TGGTCGTGTG	AAGGATGTAA	GGCCTTTTTT	360
20	AAAAGAAGCA	TTCAAGGACA	TAATGATTAT	ATTTGTCCAG	CTACAAATCA	GTGTACAATC	420
	GATAAAAACC	GGCGCAAGÀG	CTGCCAGGCC	TGCCGACTTC	GGAAGTGTTA	CGAAGTGGGA	480
25	ATGGTGAAGT	GTGGCTCCCG	GAGAGAGAGA	TGTGGGTACC	GCCTTGTGCG	GAGACAGAGA	540
30	AGTGCCGACG	AGCAGCTGCA	CTGTGCCGGĊ	AAGGCCAAGA	GAAGTGGCGG	CCACGCGCCC	600
	CGAGTGCGGG	AGCTGCTGCT	GGACGCCCTG	AGCCCCGAGC	AGCTAGTGCT	CACCCTCCTG	660
35	GAGGCTGAGC	CGCCCCATGT	GCTGATCAGC	CGCCCCAGTG	CGCCCTTCAC	CGAGGCCTCC	720
	ATGATGATGT	CCCTGACCAA	GTTGGCCGAC	AAGGAGTTGG	TACACATGAT	CAGCTGGGCC	780
40	AAGAAGATTC	CCGGCTTTGT	GGAGCTCAGC	CTGTTCGACC	AAGTGCGGCT	CTTGGAGAGC	840
45	TGTTGGATGG	AGGTGTTAAT	GATGGGGCTG	ATGTGGCGCT	CAATTGACCA	CCCCGGCAAG	900
45	CTCATCTTTG	CTCCAGATCT	TGTTCTGGAC	AGGGATGAGG	GGAAATGCGT	AGAAGGAATT	960
50	CTGGAAATCT	TTGACATGCT	CCTGGCAACT	ACTTCAAGGT	TTCGAGAGTT	AAAACTCCAA	1020 ;
	CACAAAGAAT	ATCTCTGTGT	CAAGGCCATG	ATCCTGCTCA	ATTCCAGTAT	GTACCCTCTG	1080
55	GTCACAGCGA	CCCAGGATGC	TGACAGCAGC	CGGAAGCTGG	CTCACTTGCT	GAACGCCGTG	1140

	ACCGATGCTT	TGGTTTGGGT	GATTGCCAAG	AGCGGCATCT	CCTCCCAGCA	GCAATCCATG	1200
5	CGCCTGGCTA	ACCTCCTGAT	GCTCCTGTCC	CACGTCAGGC	ATGCGAGTAA	CAAGGGCATG	1260
	GAACATCTGC	TCAACATGAA	GTGCAAAAAT	GTGGTCCCAG	TGTATGACCT	GCTGCTGGAG	1320
10	ATGCTGAATG	CCCACGTGCT	TCGCGGGTGC	AAGTCCTCCA	TCACGGGGTC	CGAGTGCAGC	1380
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20	(B) TYF (C) STI	NGTH: 1251 base PE: nucleic acid RANDEDNESS: POLOGY: linear	•				
25	(ii) MOLEC	ULE TYPE: cDN	A				
	(xi) SEQUE	NCE DESCRIPT	TION: SEQ ID NO	D: 2:			
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45	GATTACGCAT	CGGGATAȚEA	CTATGGAGTC	TGGTCGTGTG	AAGGATGTAA	GGCCTTTTTT	*360
	AAAAGAAGCA	TTCAAGGACA	TAATGATTAT	ATTTGTCCAG	CTACAAATCA	GTGTACAATC	420
50	GATAAAAACC	GGCGCAAGAG	CTGCCAGGCC	TGCCGACTTC	GGAAGTGTTA	CGAAGTGGGA	480

	ATGGTGAAGT	GTGGCTCCCG	GAGAGAGA	TGTGGGTACC	GCCTTGTGCG	GAGACAGAGA	540
5	AGTGCCGACG	AGCAGCTGCA	CTGTGCCGGC	AAGGCCAAGA	GAAGTGGCGG	CCACGCGCCC	600
40	CGAGTGCGGG	AGCTGCTGCT	GGACGCCCTG	AGCCCCGAGC	AGCTAGTGCT	CACCCTCCTG	660
10	GAGGCTGAGC	CGCCCCATGT	GCTGATCAGC	CGCCCCAGTG	CGCCCTTCAC	CGAGGCCTCC	720
15	ATGATGATGT	CCCTGACCAA	GTTGGCCGAC	AAGGAGTTGG	TACACATGAT	CAGCTGGGCC	780
	AAGAAGATTC	CCGGCTTTGT	GGAGCTCAGC	CTGTTCGACC	AAGTGCGGCT	CTTGGAGAGC	840
20	TGTTGGATGG	AGGTGTTAAT	GATGGGGCTG	ATGTGGCGCT	CAATTGACCA	CCCCGGCAAG	900
	CTCATCTTTG	CTCCAGATCT	TGTTCTGGAC	AGGGATGAGG	GGAAATGCGT	AGAAGGAATT	960
25	CTGGAAATCT	TTGACATGCT	CCTGGCAACT	ACTTCAAGGT	TTCGAGAGTT	AAAACTCCAA	1020
30	CACAAAGAAT	ATCTCTGTGT	CAAGGCCATG	ATCCTGCTCA	ATTCCAGTAT	GTACCCTCTG	1080
		·			•	GAACGCCGTG	1140
35						GCAATCCATG	1200
	CGCCTGGCTA	ACCTCCTGAT	GCTCCTGTCC	CACGTCAGGC	ATGCGAGGTG	A	1251

- 40 (2) INFORMATION FOR SEQ ID NO: 3:
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    - (A) LENGTH: 66 amino acids (B) TYPE: amino acid

    - (C) STRANDEDNESS: single
    - (D) TOPOLOGY: linear
  - (ii) MOLECULE TYPE: peptide
  - (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 3:

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5		Cys 1	Ala	Val	Cys	Ser 5	Asp	Tyr	ALa		10	туг	HIS	Tyr	GIÀ	15	Tr
		Ser	Cys	Glu	Gly 20	Cys	Lys	Ala	Phe	Phe 25	Lys	Arg	Ser	Ile	Gln 30	Gly	His
10		Asn	Asp	Tyr 35	Ile	Cys	Pro	Ala	Thr	Asn	Gln	Cys	Thr	Ile 45	Asp	Lys	Ası
15		Arg	Arg 50	Lys	Ser	Cys	Gln	Ala 55	Cys	Arg	Leu	Arg	Lys 60	Cys	Tyr	Glu	Val
20		Gly 65	Met														
25	(2) INFOR						, 3:										
30	(B)	) TYPE ) STR/	GTH: 2 E: amir ANDEI OLOG	no aci DNES	d S: sin												
35	(ii) MO (xi) SE				•	N: SE	Q ID N	NO: 4:									
40		Leu 1	Val	Leu	Thr	Leu 5	Leu	Glu	Ala	Glu	Pro 10	Pro	His	Val	Leu	Ile 15	Ser
45		Arg	Pro	Ser	Ala 20	Pro	Phe	Thr	Glu	Ala 25	Ser	Mét	Met	Met	Ser 30	Leu	Thr
		Lys	Leu	Ala 35	Asp	Lys	Glu	Leu	Val 40	His	Met ,	Ile	Ser	Trp 45	Ala	Lys	Lys
50																	
55																	

		Ile	Pro 50	Gly	Phe	Val	Glu	Leu 55	Ser	Leu	Phe	Asp	Gln 60	Val	Ärg	Leu	Leu
5		Glu 65	Ser	Cys	Trp	Met	Glu 70	Val	Leu	Met	Met	Gly 75	Leu	Met	Trp	Arg	Ser 80
10		Ile	Asp	His	Pro	Gly 85	Lys	Leu	Ile	Phe	Ala 90	Pro	Asp	Leu	Val	Leu 95	Asp
15		Arg	Asp	Glu	Gly 100	Lys •	Суѕ	Val	Glu	Gly 105	Ile	Leu	Glu	Ile	Phe 110	Asp	Met
20		Leu	Leu	Ala 115	Thr	Thr <sub>.</sub>	Ser	Arg	Phe 120	Arg	Glu	Leu	Lys	Leu 125	Gln	His	Lys
25		Glu	Tyr 130	Leu	Cys	Val	Lys	Ala 135	Met	Ile	Leu	Leu	Asn 140		Ser	Met	Tyr
30		Pro 145	Leu	Val	Ťhr	Ala	Thr 150	Gln	Asp	Ala	-	Ser 155	Ser	Arg	Lys	Leu	Ala 160
35		His	Leu	Leu	Asn	Ala 165	Val	Thr	Asp	Ala	Leu 170	Val	Trp	Val	Ile	Ala 175	Lys
		Ser	Gly	Ile	Ser 180	Ser	Gln	Gln	Gln	Ser 185	Met	Arg	Leu	Ala	Asn 190	Leu	Leu
40		Met	Leu	Leu 195	Ser	His	Val	Arg	His 200	Ala	Ser	Asn	Lys	Gly 205	Met	Glu	His
45		Leu	Leu 210	Asn	Met	Lys	Cys	Lys 215	Asn	Val	Val	Pro	Val 220	Tyr	Asp	Leu	Leu
50		Leu 225	Glu	Met	Leu	Asn	Ala 230	His	Val	Leu							•
	(2) INFORM	ATION	FOR	SEQ	ID NO	: 5:											
55	(i) SEQU	ENCE	CHA	RACT	ERIS	TICS:											
		ENGT		7 ami acid	no aci	ds											

Ç

(C) STRANDEDNESS: single (D) TOPOLOGY: unknown

_	(ii) M	OLECU	JLE T	YPE: p	roteir	1											
5	(xi) S	EQUE	NCE [	DESC	RIPTIO	ON: SI	EQ ID	NO: 5	<b>i</b> ;								
10		Met 1	Asn	Tyr	Ser	Ile 5	Pro	Ser	Asn	Val	Thr 10	Asn	Leu	Glu	Gly	Gly. 15	Pro
15		Gly	Arg	Gln	Thr 20	Thr	Ser	Pro	Asn	Val 25	Leu	Trp	Pro	Thr	Pro 30	Gly	His
20		Leu	Ser	Pro 35	Leu	Val	Val	His	Arg 40	Gln	Leu	Ser	His	Leu 45	Tyr	Ala	Glu
45		Pro	Gln 50	Lys	Ser	Pro	Trp	<b>Cys</b> 55	Glu	Ala	Arg	Ser	Leu 60	Glu	His	Thr	Leu
25		Pro 65	Val	Asn	Arg	Glu	Thr 70	Leu	Lys	Arg	Lys	Va1 75	Ser	Gly	Asn	Arg	Cys 80
30		Ala	Ser	Pro	Val	Thr 85		Pro	Gly	Ser	Lys 90	Arg	Asp	Ala	His	Phe 95	Cys
35		Ala	Val	Cys	Ser 100		Tyr	Ala	Ser	Gly 105		His	Tyr	Gly	Val 110	Trp	Ser
40		Cys	Glu	Gly 115		Lys	Ala	Phe	Phe 120		Arg	Ser	Ile	Gln 125		His	Asn
45		Asp	Tyr 130	Ile	Cys	Pro	Ala	Thr 135		Gln	Cys	Thr	11e		Lys	Asn	Arç
50																	
55																	

	Arg 145	Lys	Ser	Cys	Gln	Ala 150	Cys	Arg	Leu	Arg	Lys 155	Cys	Tyr	Glu	Val	Gly 160
5	Met	Val	Lys	Cys	Gly 165	Ser	Arg	Arg	Glu	Arg 170	Cys	Gly	Tyr	Arg	Leu 175	Val
10	Arg	Arg	Gln	Arg 180	Ser	Ala	Asp	Glu	Gln 185	Leu	His	Cys	Ala	Gly 190	Lys	Ala
15	Lys	Arg	Ser 195	Gly	Gly	His	Ala	Pro 200	Arg	Val	Arg	Glu	Leu 205	Leu	Leu	Asp
20	Ala	Leu 210	Ser	Pro	Glu	Gln	Leu 215	Val	Leu	Thr	Leu	Leu 220	Glu	Ala	Glu	Pro
25	225	His		,		230					235					240
30		Met			245					250					255	
35		Ser		260					265					270		
	ASP	GIN	275	Arg	Leu	Leu.	GIU	280	Cys	irb	Mec	GIU	285	Tea	Mec	nec
40	Gly	Leu 290	Met	Trp	Arg	Ser	11e 295	Asp	His	Pro	Gly	Lys 300	Leu	Ile	Phe	Ala
45	Pro 305	Asp	Leu	Val	Leu	Asp 310	Arg	Asp	Glu	Gly	Lys 315	Суѕ	Val	Glu	Gly	11e 320
50		Glu			325					330					335	
55	Leu	Lys	Leu	Gln 340	His	Lys	Glu	Tyr	Leu 345	Cys	Val	Lys	Ala	<b>Met</b> 350	Ile	Leu

	1	Leu	Asn	Ser 355	Ser	Met	Tyr	Pro	Leu 360	Val	Thr	Ala	Thr	Gln 365	Asp	Ala	Asp
5		Ser	ser 370	Arg	Lys	Leu	Ala	His 375	Leu	Leu	Asn	Ala	Val 380	Thr	Asp	Ala	Leu
10		Val 385	Trp	Val	Ile	Àla	Lys 390	Ser	Gly	Ile	Ser	Ser 395	Gln	Gln	Gln	Ser	Met
15	2	Arg	Leu	Ala	Asn	Leu 405	Leu	Met	Leu	Leu	Ser 410	His	Val	Arg	His	Ala 415	Ser
20	,	Asn	Lys	Gly	Met 420	Glu	His	Leu	Leu	<b>Asn</b> 425	Met	Lys	Cys	Lys	Asn 430	Val	Val
25	I	Pro	Val	Tyr 435	Asp	Leu	Leu	Leu	Glu 440	Met	Leu	Asn	Ala	His 445	Val	Leu	Arg
30	·	Sly	Cys 450	Lys	Ser	Ser	Ile	Th <i>r</i> 455	Gly	Ser	Glu	Cys	Ser 460	Pro	Ala	Glu	Asp
		Ser 465	Lys	Ser	Lys	Glu	Gly 470	Ser	Gln	Asn	Pro	Gln 475	Ser	Gln			
35	(2) INFORM	ATIO	N FOI	R SEC	N DI	O: 6:											
	(i) SEQU	JENC	CE CH	IARAC	TER	STICS	<b>3</b> :										
40	(B) <sup>-</sup> (C) <sup>3</sup>	TYPE STRA	E: amii ANDEI	116 an no aci DNES Y: unk	d S: sin												
45	(ii) MOLI	ECU	E TY	PE: pi	rotein												
	(xi) SEQ	UEN	CE DI	ESCR	IPTIO	N: SE	Q ID N	10: 6:									
50		Met	Asn	Tyr	Ser	Ile	Pro	Ser	Asn	Val	Thr	Asn	Leu	<b>Gl</b> u	Gly	Gly	Pro

	1				5					10					15	
5	Gly	Arg	Gln	Thr 20	Thr	Ser	Pro	Asn	Val 25	Leu	Trp	Pro	Thr	Pro 30	Gly	His
10	Leu	Ser	Pro 35	Leu	Val	Val	His	Arg 40	Gln	Leu	Ser	His	Leu 45	Tyr	Ala	Glu
15	Pro	Gln 50	Lys	Ser	Pro	Trp	Cys 55	Glu	Ala	Arg	Ser	Leu 60	Glu	His	Thr	Leu
20	Pro 65	Val	Asn	Arg	Glu	Thr 70	Leu	Lys	Arg	Lys	<b>Val</b> 75	Ser	Gly	Asn	Arg	Cys 80
25	Ala	Ser	Pro	Val	Thr 85	Gly	Pro	Gly	Ser	Lys 90	Arg	Asp	Ala	His	Phe 95	Cys
25			Cys	100					105					110		•
30	Cys	Glu	Gly 115	Cys	Lys	Ala	Phe	Phe 120	Lys	Arg	Ser	Ile	Gln 125	Gly	His	Asn
35	Asp	Tyr 130	Ile	Cys	Pro	Ala	Thr 135	Asn	Gln	Суз	Thr	11e 140	Asp	Lys	Asn	Arg
40	Arg 145	Lys	Ser	Cys	Gln	Ala 150	Суз	Arg	Leu	Arg	Lys 155		Tyr	Glu	Val	Gly 160
45	Met	Val	Lys	Cys	Gly 165	Ser	Arg	Arg	Glu	Arg 170	Cys	Gly	Tyr	Arg	Leu 175	Val
50	Arg	Arg	Gln	Arg 180	Ser	Ala	Asp	Glu	Gln 185	Leu	His	Cys	Ala	Gly 190	Lys	Ala
	Lys	Arg	Ser 195	Gly	Gly	His	Ala	Pro 200	Arg	Val	Arg	Glu	Leu 205	Leu	Leu	Asp
55	Ala	Leu	Ser	Pro	Glu	Gln	Leu	Val	Leu	Thr	Leu	Leu	Glu	Ala	Glu	Pro

		210					215					220				
	Pro 225	His	Val	Leu	Ile	Ser 230	Arg	Pro	Ser	Ala	Pro 235		Thr	Glu	Ala	Ser 240
10	Met	Met	Met	Ser	Leu 245	Thr	Lys	Leu	Ala	Asp 250	Lys	Glu	Leu	Val	His 255	Met
15	Ile	Ser	Trp	Ala 260	Lys	Lys	Ile	Pro	Gly 265	Phe	Val	Glu	Leu	Ser 270	Leu	Phe
20	Asp	Gln	Val 275	Arg	Leu	Leu	Glu	Ser 280	Cys	Trp	Met	Glu	Val 285	Leu	Met	Met
25	_	Leu 290	Met	Trp	Arg	Ser	Ile 295	Asp	His	Pro	Gly	Lys 300	Leu	Ile	Phe	Ala
	Pro 305	Asp	Leu	Val-	Ļeu	Asp 310	Arg	Asp	Glu	Gly	Lys 315	Cys	Val	Glu	Gly	Ile 320
30	Leu	Glu	Ile	Phe	Asp 325	Met	Leu	Leu	Ala	Thr 330	Thr	Ser	Arg	Phe	Arg 335	Glu-
35	Leu	Lys	Leu	Gln 340	His	Lys	Glu	Tyr	Leu 345	Cys	Val	Lys	Ala	Met 350	Ile	Leu
40	Leu	Asn	Ser 355	Ser	Met	Tyr	Pro	Leu 360	Val	Thr	Ala	Thr	Gln 365	Asp	Ala	Asp
45	5er	Ser 370	Arg	Lys	Leu	Ala	ніs 375	Leu	Leu	Asn	Ala	Val 380	Thr	Asp	Ala	Leu
	Val 385	Trp	<b>Val</b>	Ile	Ala	Lys 390	Ser	Gly	Ile	Ser	Se <i>r</i> 395	Gln	Gln	Gln	Ser	Met 400
55	Arg	Leu	Ala	Asn	Leu 405	Leu	Met	Leu	Leu	Ser 410	His	Val	Arg	His	Ala 415	Arg

(2) INFORMATION FOR SEQ ID NO: 7:

	(I) SEQUENCE CHARACTERISTICS:	
5	<ul><li>(A) LENGTH: 29 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: both</li><li>(D) TOPOLOGY: unknown</li></ul>	
	(ii) MOLECULE TYPE: cDNA	
10	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 7:	
15	GGIGAYGARG CWTCIGGITG YCAYTAYGG	29
	(2) INFORMATION FOR SEQ ID NO: 8:	
20	(i) SEQUENCE CHARACTERISTICS:	
	<ul><li>(A) LENGTH: 29 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
25	(ii) MOLECULE TYPE: cDNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 8:	
30	·	
	AAGCCTGGSA YICKYTTIGC CCAIYTIAT	29
35	(2) INFORMATION FOR SEQ ID NO: 9:	
	(i) SEQUENCE CHARACTERISTICS:	
40	<ul><li>(A) LENGTH: 22 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
45	(ii) MOLECULE TYPE: cDNA	
75	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 9:	
50	TGTTACGAAG TGGGAATGGT GA	22
	(2) INFORMATION FOR SEQ ID NO: 10:	
55	(i) SEQUENCE CHARACTERISTICS:	
	(A) LENGTH: 24 base pairs	

	(C) STRANDEDNESS: single (D) TOPOLOGY: linear	
_	(ii) MOLECULE TYPE: cDNA	
5	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 10:	
10	TTGACACCAG ACCAACTGGT AATG	24
	(2) INFORMATION FOR SEQ ID NO: 11:	
15	(i) SEQUENCE CHARACTERISTICS:	
20	<ul><li>(A) LENGTH: 24 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
	(ii) MOLECULE TYPE: cDNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 11:	
25	GGTGGCGACG ACTCCTGGAG CCCG	. 24
30	(2) INFORMATION FOR SEQ ID NO: 12:	
	(i) SEQUENCE CHARACTERISTICS:	
35	<ul><li>(A) LENGTH: 22 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
	(ii) MOLECULE TYPE: cDNA	
40	(XI) SEQUENCE DESCRIPTION: SEQ ID NO: 12:	
	GTACACTGAT TTGTAGCTGG AC	22
45	(2) INFORMATION FOR SEQ ID NO: 13:	
	(i) SEQUENCE CHARACTERISTICS:	
50	<ul><li>(A) LENGTH: 20 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
55	(ii) MOLECULE TYPE: cDNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 13:	

	CCATGATGAT GTCCCTGACC	20
5	(2) INFORMATION FOR SEQ ID NO: 14:	
	(i) SEQUENCE CHARACTERISTICS:	
10	<ul><li>(A) LENGTH: 20 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
15	(ii) MOLECULE TYPE: cDNA	
13	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 14:	
20	TCGCATGCCT GACGTGGGAC	20
	(2) INFORMATION FOR SEQ ID NO: 15:	
25	(i) SEQUENCE CHARACTERISTICS:	
	<ul><li>(A) LENGTH: 24 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
30	(ii) MOLECULE TYPE: cDNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 15:	
35		
	GGCSTCCAGC ATCTCCAGSA RCAG	24
40	(2) INFORMATION FOR SEQ ID NO: 16:	
	(i) SEQUENCE CHARACTERISTICS:	
45	<ul><li>(A) LENGTH: 20 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
50	(ii) MOLECULE TYPE: cDNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 16:	
55	GGAAGCTGGC TCACTTGCTG	20
	(2) INFORMATION FOR SEQ ID NO: 17:	

	(i) SEQUENCE CHARACTERISTICS:	
5	<ul><li>(A) LENGTH: 20 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
	(ii) MOLECULE TYPE: cDNA	
10	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 17:	
	TCTTGTTCTG GACAGGGATG	20
15	(2) INFORMATION FOR SEQ ID NO: 18:	
	(i) SEQUENCE CHARACTERISTICS:	
20	<ul><li>(A) LENGTH: 20 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
25	(ii) MOLECULE TYPE: cDNA	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 18:	
30	GCATGGAACA TCTGCTCAAC	20
	(2) INFORMATION FOR SEQ ID NO: 19:	
35	(i) SEQUENCE CHARACTERISTICS:	
40	<ul><li>(A) LENGTH: 21 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
	(ii) MOLECULE TYPE: cDNA	
45	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 19:	
	AGCAAGTTCA GCCTGTTAAG T	. 21
50	(2) INFORMATION FOR SEQ ID NO: 20:	
	(i) SEQUENCE CHARACTERISTICS:	
55	(A) LENGTH: 1257 base pairs (B) TYPE: nucleic acid (C) STRANDEDNESS: double (D) TOPOLOGY: linear	

(ii) M	IOLECUI	LE TYPE:	cDNA
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(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 20:

: 60
: 120
. 180
240
300
360
c

	AAAAGAAGCA	TTCAAGGACA	TAATGATTAT	ATTTGTCCAG	CTACAAATCA	GTGTACAATC	420
5	GATAAAAACC	GGCGCAAGAG	CTGCCAGGCC	TGCCGACTTC	GGAAGTGTTA	CGAAGTGGGA	480
	ATGGTGAAGT	GTGGCTCCCG	GAGAGAGAGA	TGTGGGTACC	GCCTTGTGCG	GAGACAGAGA	540
10	AGTGCCGACG	AGCAGCTGCA	CTGTGCCGGC	AAGGCCAAGA	GAAGTGGCGG	CCACGCGCCC	600
45	CGAGTGCGGG	AGCTGCTGCT	GGACGCCCTG	AGCCCCGAGC	AGCTAGTGCT	CACCCTCCTG	660
15	GAGGCTGAGC	CGCCCCATGT	GCTGATCAGC	CGCCCCAGTG	CGCCCTTCAC	CGAGGCCTCC	720
20	ATGATGATGT	CCCTGACCAA	GTTGGCCGAC	AAGGAGTTGG	TACACATGAT	CAGCTGGGCC	780
	AAGAAGATTC	CCGGCTTTGT	GGAGCTCAGC	CTGTTCGACC	AAGTGCGGCT	CTTGGAGAGC	840
?5	TGTTGGATGG	AGGTGTTAAT	GATGGGGCTG	ATGTGGCGCT	CAATTGACCA	CCCCGGCAAG	900
	CTCATCTTTG	CTCCAGATCT	TGTTCTGGAC	AGGGATGAGG	GGAAATGCGT	AGAAGGAATT	960
30	CTGGAAATCT	TTGACATGCT	CCTGGCAACT	ACTTCAAGGT	TTCGAGAGTT	AAAACTCCAA	1020
	CACAAAGAAT	ATCTCTGTGT	CAAGGCCATG	ATCCTGCTCA	ATTCCAGTAT	GTACCCTCTG	1080
35	GTCACAGCGA	CCCAGGATGC	TGACAGCAGC	CGGAAGCTGG	CTCACTTGCT	GAACGCCGTG	1140
o	ACCGATGCTT	TGGTTTGGGT	GATTGCCAAG	AGCGGCATCT	CCTCCCAGCA	GCAATCCATG	1200
	CGCCTGGCTA	ACCTCCTGAT	GCTCCTGTCC	CACGTCAGGC	ATGCGAGGTC	TGCCTGA	1257

(2) INFORMATION FOR SEQ ID NO: 21:

45

50

- (i) SEQUENCE CHARACTERISTICS:
  - (A) LENGTH: 418 amino acids
  - (B) TYPE: amino acid
  - (C) STRANDEDNESS: single
  - (D) TOPOLOGY: linear
- (ii) MOLECULE TYPE: protein
- (xi) SEQUENCE DESCRIPTION: SEQ ID NO: 21:

	Met 1	Asn	Tyr	Ser	Ile 5	Pro	Ser	Asn	Val	Thr 10	Asn	Leu	Glu	Gly	Gly 15	Pro
5	Gly	Arg	Gln	Thr 20	Thr	Ser	Pro	Asn	Val 25	Leu	Trp	Pro	Thr	Pro 30	Gly	His
10	Leu	Ser	Pro 35	Leu	Val	Val	His	Arg 40	Gln	Leu	Ser	His	Leu 45	туг	Ala	Glu
15	Pro	Gln 50	Lys	Ser	Pro	Trp	<b>Cys</b> 55	Glu	Ala	Arg	Ser	Leu 60	Glu	His	Thr	Leu
20	Pro 65	Val	Asn	Arg	Glu	Thr 70	Leu	Lys	Arg	Lys	Val 75	Ser	Gly	Asn	Arg	Cys 80
25	Ala	Ser	Pro	Val	Thr 85	Gly	Pro	Gly	Ser	Lys 90	Arg	Asp	Ala	His	Phe 95	Cys
30	Ala	Val	Cys	Ser 100	Asp	Tyr	Ala	Ser	Gly 105	Tyr	His	Tyr	Gly	Val 110	Trp	Ser
35	Суз	Glu	Gly 115	Cys	Lys	Ala	Phe	Phe 120	Lys	Arg	Ser	Ile	Gln 125	Gly	His	Asn
	Asp	Tyr 130	Ile	Cys	Pro	Ala	Thr 135	Asn	Gln	Cys	Thr	Ile 140	Asp	Lys	Asn	Arg
40	Arg 145	Lys	Ser	Cys	Gln	Ala 150	Cys	Arg	Leu	Arg	Lys 155	Суѕ	Tyr	Glu	Val	Gly 160
45	Met	Val	Lys	Cys	Gly 165	Ser	Arg	Arg	Glu	Arg 170	Суѕ	Gly	Tyr	Arg	Leu 175	Val
50	Arg	Arg	Gln	Arg	Ser	Ala	Asp	Glu	Gln	Leu	His	Cys	Ala	Gly	Lys	Ala

				180	)				18:	5				19	0	
5	Lys	a Ar	g Ser 195		/ Gly	y His	. Ala	200		g Va	l Arç	g Glu	20!		ı Le	u As <sub>i</sub>
10	Ala	210	u Ser	Pro	Glu	ı Gln	215		l Let	ı Thi	r Leu	1 Let 220		ı Ala	Gl:	u Pro
15	Pro 225		s Val	Leu	Ile	Ser 230		Pro	Ser	: Ala	235		Th:	Glu	Ala	240
20	Met	Met	: Met	Ser	Leu 245		Lys	Leu	Ala	Asp 250		Glu	Leu	Val	His 255	
25	Ile	Ser	Trp	Ala 260	Lys	Lys	Ile	Pro	Gly 265	Phe	Val	Glu	Leu	Ser 270		Phe
	Asp	Gln	Val 275	Arg	Leu	Leu	Glu	Ser 280	Суѕ	Trp	Met	Glu	Val 285	Leu	Met	Met
30	Gly	Leu 290	Met	Trp	Arg	Ser	Ile 295	Asp	His	Pro	Gly	Lys 300	Leu	Ile	Phe	Ala
35	Pro 305	Asp	Leu	Val	Leu	Asp 310	Arg	Asp	Glu	Gly	Lys 315	Cys	Val	Glu	Gly	Ile 320
40	Leu	Glu	Ile	Phe	Asp 325	Met	Leu	Leu	Ala	Thr 330	Thr	Ser	Arg	Phe	Arg 335	Glu
45	Leu	Lys	Leu	Gln 340	His	Lys	Glu	Tyr	Leu 345	Cys	Val	Lys	Ala	<b>Met</b> 350	Ile	Leu
50	Leu	Asn	Ser 355	Ser	Met	Tyr	Pro	Leu 360	Val	Thr	Ala	Thr	Gln 365	Asp	Ala	Asp
		Ser 370	Arg	Lys	Leu		His 375	Leu	Leu	Asn		Val 380	Thr	Asp	Ala	Leu
55	Val	Trp	Val	Ile .	Ala	Lys	Ser	Gly	Ile	Ser	Ser	Gln	Gln	Gln	Ser	Met

	385		. 3	390				395					400
5	Arg	Leu Ala Asn	Leu I	Leu Met	Leu	Leu	Ser 410	His	Val	Arg	His	Ala 415	Arg
10	Ser .	Ala						. • •		,			
	(2) INFORMATION	N FOR SEQ ID N	O: <b>22</b> :										
15	(i) SEQUENC	E CHARACTERI	STICS:										
20	(B) TYPE (C) STRA	iTH: 34 base pair i: nucleic acid NDEDNESS: sin DLOGY: linear		٠.									
	(ii) MOLECUL	E TYPE: cDNA											
25	(xi) SEQUENC	CE DESCRIPTIO	N: SEQ	ID NO: 22	2:								
	CTTGGATCCA TA	GCCCTGCT GT	GATGA	ATT ACA	G								34
30	(2) INFORMATION	N FOR SEQ ID N	O: 23:				,						
	(i) SEQUENC	E CHARACTER!	STICS:										
35	(B) TYPE: (C) STRA	TH: 33 base pairs : nucleic acid NDEDNESS: sing LOGY: linear											
40	(ii) MOLECUL	E TYPE: cDNA											
70	(xi) SEQUENC	CE DESCRIPTIO	N: SEQ	ID NO: 23	:								
45	GATGGATCCT CA	CCTCAGGG CC	AGGCG1	rca ctg									33
	(2) INFORMATION	FOR SEQ ID NO	D: 24:										
50	(i) SEQUENCE	E CHARACTERIS	STICS:										
55	(B) TYPE: (C) STRAN	TH: 1898 base pa nucleic acid NDEDNESS: sing LOGY: linear											
	(ii) MOLECULE	E TYPE: cDNA											
	(xi) SEQUENC	E DESCRIPTION	I: SEQ I	D NO: 24:	:								

	CACGAATCTT	TGAGAACATT	ATAATGACCT	TTGTGCCTCT	TCTTGCAAGG	TGTTTTCTCA	. 60
5	GCTGTTATCT	CAAGACATGG	Ататааааа	CTCACCATCT	AGCCTTAATT	стссттсстс	120
	CTACAACTGC	AGTCAATCCA	TCTTACCCCT	GGAGCACGGC	TCCATATACA	TACCTTCCTC	180
10	CTATGTAGAC	AGCCACCATG	AATATCCAGC	CATGACATTC	TATAGCCCTG	CTGTGATGAA	240
	TTACAGCATT	CCCAGCAATG	TCACTAACTT	GGAAGGTGGG	CCTGGTCGGC	AGACCACAAG	300
15	CCCAAATGTG	TTGTGGCCAA	CACCTGGGCA	CCTTTCTCCT	TTAGTGGTCC	ATCGCCAGTT	360
20	ATCACATCTG	TATGCGGAAC	CTCAAAAGAG	TCCCTGGTGT	GAAGCAAGAT	CGCTAGAACA	420
	CACCTTACCT	GTAAACAGAG	AGACACTGAA	AAGGAAGGTT	AGTGGGAACC	GTTGCGCCAG	480
25	CCCTGTTACT	GGTCCAGGTT	CAAAGAGGGA	TGCTCACTTC	TGCGCTGTCT	GCAGCGATTA	540
	CGCATCGGGA	TATCACTATG	GAGTCTGGTC	GTGTGAAGGA	TGTAAGGCCT	TTTTTAAAAG	600
30	AAGCATTCAA	GGACATAATG	ATTATATTTG	TCCAGCTACA	AATCAGTGTA	CAATCGATAA	660
	AAACCGGCGC	AAGAGCTGCC	AGGCCTGCCG	ACTTCGGAAG	TGTTACGAAG	TGGGAATGGT	720

	GAAGTGTGGC	TCCCGGAGAG	AGAGATGTGG	GTACCGCCTT	GTGCGGAGAC	AGAGAAGTGC	780
5	CGACGAGCAG	CTGCACTGTG	CCGGCAAGGC	CAAGAGAAGT	GGCGGCCACG	CGCCCCGAGT	840
	GCGGGAGCTG	CTGCTGGACG	CCCTGAGCCC	CGAGCAGCTA	GTGCTCACCC	TCCTGGAGGC	90,0
10	TGAGCCGCCC	CATGTGCTGA	TCAGCCGCCC	CAGTGCGCCC	TTCACCGAGG	CCTCCATGAT	960
15	GATGTCCCTG	ACCAAGTTGG	CCGACAAGGA	GTTGGTACAC	ATGATCAGCT	GGGCCAAGAA	1020
	GATTCCCGGC	TTTGTGGAGC	TCAGCCTGTT	CGACCAAGTG	CGGCTCTTGG	AGAGCTGTTG	1080
20	GATGGAGGTG	TTAATGATGG	GGCTGATGTG	GCGCTCAATT	GACCACCCCG	GCAAGCTCAT	1140
	CTTTGCTCCA	GATCTTGTTC	TGGACAGGGA	TGAGGGGAAA	TGCGTAGAAG	GAATTCTGGA "	1200
25	AATCTTTGAC	ATGCTCCTGG	CAACTACTTC	AAGGTTTCGA	GAGTTAAAAC	TCCAACACAA	1260
30	AGAATATCTC	TGTGTCAAGG	CCATGATCCT	GCTCAATTCC	AGTATGTACC	CTCTGGTCAC	1320
30	AGCGACCCAG	GATGCTGACA	GCAGCCGGAA	GCTGGCTCAC	TTGCTGAACG	CCGTGACCGA	1380
35	TGCTTTGGTT	TGGGTGATTG	CCAAGAGCGG	CATCTCCTCC	CAGCAGCAAT	CCATGCGCCT	1440
	GGCTAACCTC	CTGATGCTCC	TGTCCCACGT	CAGGCATGCG	AGTAACAAGG	GCATGGAACA	1500
40	TCTGCTCAAC	: ATGAAGTGCA	AAAATGTGGT	CCCAGTGTAT	GACCTGCTGC	TGGAGATGCT	1560
	GAATGCCCAC	: GTGCTŢCGCG	GGTGCAAGTC	CTCCATCACG	GGGTCCGAGT	GCAGCCCGGC	1620
45	AGAGGACAGI	* AAAAGCAAAG	AGGGCTCCCA	GAACCCACAG	TCTCAGTGAC	GCCTGGCCCT	1680
50						GTCAGGAGCC	1740 •
						GGTCATGTAC	1800
EE	CATCCTTCC	C TCCACCTTCC	CAACTCTCAG	GAGTCGGTG	GAĞGAAGCCA	TAGTTTCCCT	1860

	TGTTAGC	AGA	GGGA	CATT	TG A	ATCG	AGCG	т тт	CCAC	AC							1898
5	(2) INFORM	MATIC	N FO	R SEC	א מו כ	O: 25:											
												,					
	(i) SEQ	MEIN	JE GF	IAHAC	JIERI	SHOS	<b>5</b> :		•								
10	(B) (C)	TYPI STR	GTH: 8 E: ami ANDE OLOG	no aci DNES	d S: sin												
15	(ii) MOI	LECU	LE TY	PE: p	eptide												
	(xi) SE	QUEN	ICE D	ESCR	IPTIO	N: SE	Q ID 1	NO: 25	5:							-	
20		Met	Asp	Ile	Lys	Asn	Ser	Pro	Ser	Ser	Leu	Asn	Ser	Pro	Ser	Ser	Tyr
		1				5					10					15	•
25		Asn	Cys	Ser	Gln 20	Ser	Ile	Leu	Pro	Leu 25	Glu	His	Gly	Ser	Ile 30	Tyr	Ile
30		Pro	Ser	Ser 35	Tyr	Val	Asp	Ser	His 40	His	Glu	Tyr	Pro		Met	Thr	Phe
50				33					40					45			•
		Tyr	Ser	Pro	Ala	Val	Met	Asn	Tyr	Ser	Ile	Pro	Ser	Asn	Val	Thr	Asn
35	,		50					55					60				
		Leu	Glu	Gly	Gly	Pro	Gly	Arg	Gln	Thr	Thr	Ser	Pro	Asn	Val	Leu	Trp
		65					70					75					80
40		Pro	Thr	Pro	Glv	His	Leu	Ser	Pro	Leu	۷ə۱	Val	Wie	A = ~	C) =	T a	C
						85					90			ALY	GIII	95	Ser
45																	
		His	Leu	Tyr		Glu	Pro	Gln	Lys		Pro	Trp	Cys	Glu	Ala	Arg	Ser
					100					105					110		
50		Leu	Glu	His	Thr	Leu	Pro	Val	Asn	Arg	Glu	Thr	Leu	Lys	Arg	Lys	Val

		•	115	5				120					125	5		
5	Ser	: Gly		Arg	, Cys	Ala	Ser 135		Val	Thr	: Gly	Pro		/ Ser	Lys	a Arg
10	Asp 145		His	Phe	: Cys	Ala 150		Cys	Ser	: Asp	Туг 155		Ser	: Gly	Tyr	His 160
15	Tyr	Gly	Val	Trp	Ser 165		Glu	Gly	Суѕ	Lys 170		Phe	Phe	Lys	Arg 175	Ser
20	Ile	Gln	Gly	His 180	Asn	Asp	Tyr	Ile	Cys 185	Pro	Ala	Thr	Asn	Gln 190	Cys	Thr
05	Ile	Asp	Lys 195	Asn	Arg	Arg	Lys	Ser 200	Cys	Gln	Ala	Cys	Arg 205	Leu	Arg	Lys
25	Cys	Tyr 210	Glu	Val	Gly	Met	Val 215	Lys	Cys	Gly	Ser	Arg 220	Arg	Glu	Arg	Cys
30	Gly 225	Tyr	Arg	Leu	Val	Arg 230	Arg	Gln	Arg	Ser	Ala 235	Asp	Glu	Gln	Leu	His 240
35	Cys	Ala	Gly	Lys	Ala 245	Lys	Arg	Ser	Gly	Gly 250	His	Ala	Pro		Val 255	Arg
40	Glu	Leu	Leu	Leu 260	Asp	Ala	Leu		Pro 265	Glu	Gln	Leu	Val	Leu 270	The	Leu
45	Leu	Glu	Ala 275	Glu	Pro	Pro	His	Val 280	Leu	Ile	Ser	Arg	Pro 285	Ser	Ala	Pro
50	Phe	Thr 290	Glu	Ala	Ser	Met	Met 295	Met	Ser	Leu	Thr	Lys 300	Leu	Ala	Asp	Lys
50	Glu 305	Leu	Val	His	Met	Ile 310	Ser	Trp	Ala		Lys 315	Ile	Pro	Gly	Phe	Val 320
55	Glu	Leu	Ser	Leu	Phe .	Asp	Gln	Val .	Arg	Leu	Leu	Glu	Ser	Cys	Trp	Met

•					325					330	1				335	<b>i</b>
5	G.	Lu Va	l Leu	Met 340		Gly	Leu	. Met	Trp		Ser	Ile	Asp	His 350		Gly
10	L	s Le	11e 355		Ala	Pro	Авр	Leu 360		Leu	Asp	Arg	Asp 365		Gly	Lys
15	cy	's Val		Gly	Ile	Leu	Glu 375		Phe	Asp	Met	Leu 380	Leu	Ala	Thr	Thr
	Se 38	r Arç	, Phe	Arg	Glu	Leu 390		Leu	Gln	His	Lys 395	Glu	Tyr	Leu	Cys	Val 400
20	Ly	s Ala	. Met	Ile	Leu 405	Leu	Asn	Ser	Ser	Met 410	Tyr	Pro	Leu	Val	Thr 415	Alā
25	Th	r Gln	Asp	Ala 420	Asp	Ser	Ser	Arg	Lys 425	Leu	Ala	His	Leu	Leu 430	Asn	Ala
30	Va	1 Thr	Asp 435	Ala	Leu	Val	Trp	Val 440	Ile	Ala	Lys	Ser	Gly 445	Ile	Ser	Ser
35	Gl	n Gln 450		Ser	Met	Arg	Leu 455	Ala	Asn	Leu	Leu	Met 460	Leu	Leu	Ser	His
	∨a 46	l Arg 5	His	Ala	Ser	Asn 470	Lys	Gly	Met	Glu	His 475	Leu	Leu	Asn	Met	Lys 480
40	Су	s Lys	Asn	Val	Val 485	Pro	Val	Tyr	Asp	Leu 490	Leu	Leu	Glu	Met	<b>Le</b> u 495	Asn
45	ΥT	a His	Val	<b>Leu</b> 500	Arg	Gly	Cys	Lys	Ser 505	Ser	Ile	Thr	Gly	Ser 510	Glu	Cys
50	Se	r Pro	Ala 515	Glu	Asp	Ser		Ser 520	Lys	Glu	GГу		Gln 525	Asn	Pro	Gln
55	Se	r Gln	r													

	(2) INFORMATION FOR SEQ ID NO: 26:	
	(i) SEQUENCE CHARACTERISTICS:	
5	<ul><li>(A) LENGTH: 30 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
10	(ii) MOLECULE TYPE: other nucleic acid	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 26	d:
15	GTGCGGATCC TCTCAAGACA TGGATATAAA	30
20	(2) INFORMATION FOR SEQ ID NO: 27:	
20	(i) SEQUENCE CHARACTERISTICS:	
25	<ul><li>(A) LENGTH: 25 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li><li>(D) TOPOLOGY: linear</li></ul>	
	(ii) MOLECULE TYPE: other nucleic acid	
30	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 2	<b>7</b> :
35	AGTAACAGGG CTGGCGCAAC GGTTC	25
	(2) INFORMATION FOR SEQ ID NO: 28:	
40	(i) SEQUENCE CHARACTERISTICS:	
	<ul><li>(A) LENGTH: 22 base pairs</li><li>(B) TYPE: nucleic acid</li><li>(C) STRANDEDNESS: single</li></ul>	
45	(D) TOPOLOGY: linear	
	(ii) MOLECULE TYPE: other nucleic acid	
	(xi) SEQUENCE DESCRIPTION: SEQ ID NO: 20	3:
50	ACTGGCGATG GACCACTAAA GG	. 22

### 55 Claims

1. Isolated estrogen receptor having an N-terminal domain, a DNA-binding domain, and a ligand-binding domain, wherein the amino acid sequence of said DNA-binding domain exhibits at least 80% homology with the amino acid

sequence shown in SEQ ID NO:3 and the amino acid sequence of said ligand-binding domain of said estrogen receptor exhibits at least 70 % homology with the amino acid sequence shown in SEQ ID NO:4, provided that the estrogen receptor does not have the amino acid sequence:

	MTFYS	PAVMN	YSVPG	STSNL	DGGPV	RLSTS	PNVLW	PTSGH	LSPLA
	THCQS	SLLYA	EPQKS	PWCEA	RSLEH	TLPVN	RETLK	RKLSG	SSCAS
	PVTSP	NAKRD	AHFCP	VCSDY	ASGYH	YGVWS	CEGCK	AFFKR	SIQGH
10	NDYIC	PATNQ	CTIDK	NRRKS	CQACR	LRKCY	EVGMV	KCGSR	RERCG
	YRIVR	RQRSS	SEQVH	CLSKA	KRNGG	HAPRV	KELLL	STLSP	EQLVL
	TLLEA	EPPNV	LVSRP	SMPFT	EASMM	MSLTK	LADKE	LVHMI	GWAKK
15	IPGFV	ELSLL	DQVRL	LESCW	MEVLM	VGLMW	RSIDH	PGKLI	FAPDL
	VLDRD	EGKCV	EGILE	IFDML	LATTS	RFREL	KLQHK	EYLCV	KAMIL
	LNSSM	YPLAS	ANQEA	ESSRK	LTHLL	NAVTD	ALVWV	IAKSG	ISSQQ
20	QSVRL	ANLLM	LLSHV	RHISN	KGMEH	LLSMK	CKNVV	PVYDL	LLEML
	NAHTL	RGYKS	SISGS	ECSST	EDSKN	KESSQ	NLQSQ		

- 2. Isolated estrogen receptor according to claim 1, **characterised in that** the amino acid sequence of said DNA-binding domain exhibits at least 90% homology with the amino acid sequence shown in SEQ ID NO: 3.
  - 3. Isolated estrogen receptor according to anyone of claims 1-2, **characterised in that** the amino acid sequence of said ligand-binding domain exhibits at least 75% homology with the amino acid sequence shown in SEQ ID NO: 4.
- Isolated estrogen receptor according to anyone of claims 1-3, characterised in that said estrogen receptor comprises the amino acid sequence of SEQ ID NO: 5, SEQ ID NO: 6, SEQ ID NO: 21 or SEQ ID NO: 25.
  - 5. Isolated DNA encoding an estrogen receptor according to claims 1-4.

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- Isolated DNA according to claim 5, characterised in that said DNA comprises the nucleic acid sequence of SEQ ID NO: 1, SEQ ID NO:2, SEQ ID NO:20 or SEQ ID NO:24.
  - 7. A recombinant expression vector comprising the DNA according to claim 5 or 6.
- 40 8. A cell transfected with DNA according to claim 5 or 6 or an expression vector according to claim 7.
  - 9. A cell according to claim 8 which is a stable transfected cell line which expresses the estrogen receptor according to any of the claims 1-4.
- 45 10. Use of a DNA according to claim 5 or 6, an expression vector according to claim 7, a cell according to claim 8 or 9, a receptor according to any one of claims 1-4, in a screening assay for identification of new drugs.
  - Method of identifying functional ligands for a receptor according to any one of claims 1-4, said method comprising the steps of
    - a) introducing into a suitable host cell 1) DNA according to claims 5 or 6, and 2) a suitable reporter gene functionally linked to an operative hormone responsive element (HRE), said HRE being able to be activated by the DNA-binding domain of the receptor encoded by said DNA;
    - b) bringing said host cell from a) into contact with potential ligands which will possibly bind to the ligand-binding domain of the receptor encoded by said DNA from step a); and
    - c) monitoring the expression of the receptor encoded by said reporter gene of step a).

#### Patentansprüche

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1. Isolierter Östrogenrezeptor mit einer N-terminalen Domäne, eine DNA-Bindungsdomäne und einer Ligandenbindungsdomäne, worin die Aminosäuresequenz der genannten DNA-Bindungsdomäne mindestens 80% Homologie mit der in SEQ ID Nr.:3 gezeigten Aminosäuresequenz besitzt und die Aminosäuresequenz der genannten Ligandenbindungsdomäne des genannten Östrogenrezeptors mindestens 70% Homologie mit der in SEQ ID Nr.:4 gezeigten Aminosäuresequenz aufweist, vorausgesetzt, dass der Östrogenrezeptor nicht die folgenden Aminosäuresequenz besitzt:

	MTFY	S PAVMN	YSVPG	STSNL	DGGPV	RLSTS	PNVLW	PTSGH	LSPLA
	THCQ	S SLLYA	EPQKS	PWCEA	RSLEH	TLPVN	RETLK	RKLSG	SSCAS
	PVTS	P NAKRD	AHFCP	VCSDY	ASGYH	YGVWS	CEGCK	AFFKR	SIQGH
15	NDYI	C PATNO	CŤIDK	NŘŘKS	CQACR	LRKCY	EVGMV	KCGSR	RERCG
	YRIV	r RQRSS	SEQVH	CLSKA	KRNGG	HAPRV	KELLL	STLSP	EQLVL
	TLLE	A ÉPPNV	LVSRP	SMPFT	EASMM	MSLTK	LADKE	<b>LAHWI</b>	GWAKK
20	IPGF	V ELSLL	DQVRL	LESCW	MEVLM	VGLMW	RSIDH	PGKLI	FAPDL
	VLDR	D EGKÇV	EGILE	IFDML	LATTS	RFREL	KLQHK	EYLCV	$\textbf{KAMIL}_{\!$
	LNSSI	M YPLAS	ANQEA	ESSRK	LTHLL	NAVTD	ALVWV.	IAKSG	issQQ
25	QSVR.	L ANLLM	LLSHV	RHISN	KGMEH	LLSMK	CKNVV	PAADT	LLEML
	, NАНТ	L RGYKŞ	SISGS	ECSST	EDSKN	KESSQ	NLQSQ		٠

- Isolierter Östrogenrezeptor nach Anspruch 1, dadurch gekennzeichnet, dass die Aminosäuresequenz der genannten DNA-Bindungsdomäne mindestens 90% Homologie mit der in der SEQ ID. Nr.:3 gezeigten Aminosäuresequenz hat.
  - isolierter Östrogenrezeptor nach einem der Ansprüche 1-2, dadurch gekennzeichnet, dass die Aminosäuresequenz der genannten Ligandenbindungsdomäne mindestens 75% Homologie mit der in der SEQ ID. Nr.:4 gezeigten Aminosäuresequenz hat.
    - Isolierter Östrogenrezeptor nach einem der Ansprüche 1-3, dadurch gekennzelchnet, dass der genannte Östrogenrezeptor die Aminosäuresequenz der SEQ ID Nr.:5, SEQ ID Nr.:6, SEQ ID Nr.:21 oder SEQ ID Nr.:25 umfasst.
- Isolierte DNA, welche für einen Östrogenrezeptor nach den Ansprüchen 1-4 codiert.
  - Isolierte DNA nach Anspruch 5, dadurch gekennzeichnet, dass die genannte DNA die Nukleinsäuresequenz der SEQ ID Nr.:1, SEQ ID Nr.:2, SEQ ID Nr.:20 oder SEQ ID Nr.:24 umfasst.
- 7. Rekombinanter Expressionsvektor, welcher die DNA nach einem der Ansprüche 5 oder 6 umfasst.
  - Zelle, welche mit der DNA nach einem der Ansprüche 5 oder 6 oder einem Expressionsvektor nach Anspruch 7 transfiziert ist.
- Zelle nach Anspruch 8, welche eine stabil transfizierte Zelllinie ist, die den Östrogenrezeptor nach einem der Ansprüche 1-4 exprimiert.
- 10. Verwendung einer DNA nach einem der Ansprüche 5 oder 6, eines Expressionsvektors nach Anspruch 7, einer Zelle nach Anspruch 8 oder 9, einem Rezeptor nach einem der Ansprüche 1-4 in einem Screening-Assay zur Identifizierung neuer Arzneimittel.
  - 11. Verfahren zur Identifizierung funktionaler Liganden für einen Rezeptor nach einem der Ansprüche 1-4, worin das genannte Verfahren die folgenden Schritte umfasst:

- a) Einbringen in eine geeignete Wirtszelle von 1) DNA nach den Ansprüchen 5 dder 6 und 2) von einem geeigneten Reporter-Gen, das funktional mit einem operativen responsiven Hormonelement (HRE) verbunden ist, wobei das genannte HRE fähig ist, von der DNA-Bindungsdomäne des von der genannten DNA codierten Rezeptors aktiviert zu werden;
- b) in Kontakt Bringen der genannten Wirtszelle von a) mit potentiellen Liganden, die möglicherweise an die Ligandenbindungsdomäne des von der genannten DNA codierten Rezeptors von Schritt a) binden und
- c) Überwachen der Expression des Rezeptors, der von dem genannten Reportergen von Schritt a) codiert wird.

#### 10 Revendications

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1. Récepteur d'oestrogène isolé ayant un domaine N-terminal, un domaine de liaison d'ADN, et un domaine de liaison de ligand, dans lequel la séquence d'acides aminés dudit domaine de liaison d'ADN présente au moins 80 % d'homologie avec la séquence d'acides aminés indiquéc dans SEQ ID N°3 et la séquence d'acides aminés dudit domaine de liaison de ligand dudit récepteur d'oestrogène présente au moins 70 % d'homologic avec la séquence d'acides aminés indiquée dans SEQ ID N°4.

à condition que le récepteur d'oestrogène ne possède pas la séquence d'acides aminés:

20	MTFYS	PAVMN	Y\$VPG	STSNL	DGGPV	RLSTS	PNVLW	PTSGH	I.SPLA
	THCQS	SLLYA	<b>EPQKS</b>	PWCEA	RSLEH	TLPVN	RETLK	RKLSG	SSCAS
	PVTSP	NAKRD	AHFCP	VCSDY	ASGYH	YGVWS	CEGCK	AFFKR	SIQGH
25	NDYIC	PATNQ	CJ.IDK	NRRKS	CQACR	LRKCY	EVGMV	KCGSR	RERCG
25	YRIVR	RQRSS	SEQVH	CLSKA	KRNGG	HAPRV	KELLL	STLSP	EQLVL
	TLLEA	EPPNV	LVSRP	SMPFT	EASMM	MSLTK	LADKE	LVHMI	GWAKK
	IPGFV	ELSLL	DQVRL	LESCW	MEVLM	VGLMW	RSIDH	PGKLI	FAPDL
30	VLDRD	EGKCV	<b>FGILE</b>	IFDML	LATTS	RFREL	KI.QHK	EYLCV	KAMII.
	LNSSM	YPLAS	$ANQE \Lambda$	ESSRK	LTHILL	NAVTD	ALVWV	LAKSG	ISSQQ
	QSVRL	ANLLM	LLSHV	RHISN	KGMEH	LLSMK	CKNVV	PVYDL	I.LEML
35	NAHTL	RGYKS	SISGS	ECSST	EDSKN	KESSQ	NLQSQ		

 Récepteur d'oestrogène isolé selon la revendication 1, caractérisé en ce que la séquence d'acides aminés dudit domaine de liaison d'ADN présente au moins 90 % d'homologie avec la séquence d'acides aminés indiquée dans SEQ ID N°3.

- 3. Récepteur d'oestrogène isolé selon l'une quelconque des revendications 1 et 2, caractérisé en ce que la séquence d'acides aminés dudit domaine de liaison de ligand présente au moins 75 % d'homologie avec la séquence d'acides aminés indiquée dans SEQ ID N°4.
- 45 4. Récepteur d'oestrogène isolé selon l'une quelconque des revendications 1 à 3, caractérisé en ce que ledit récepteur d'oestrogène comprend une séquence d'acides aminés de SEQ ID N°5, SEQ ID N°6, SEQ ID N°21 ou SEQ ID N°25.
  - 5. ADN isolé codant pour un récepteur d'oestrogène selon l'une quelconque des revendications 1 à 4.
  - 6. ADN isolé selon la revendication 5, caractérisé en ce que ledit ADN comprend la séquence d'acide nucléique de SEQ ID N°1, SEQ ID N°2, SEQ ID N°20 ou SEQ ID N°24.
  - 7. Vecteur d'expression recombinant comprenant l'ADN selon l'une quelconque des revendications 5 ou 6.
  - 8. Cellule transfectée avec l'ADN selon l'une quelconque des revendications 5 ou 6 ou un vecteur d'expression selon la revendication 7.

- Cellule selon la revendication 8 qui est une lignée cellulaire transfectée stable qui exprime le récepteur d'oestrogène selon l'une quelconque des revendications 1 à 4.
- 10. Utilisation d'un ADN selon l'une quelconque des revendications 5 ou 6, d'un vecteur d'expression selon la revendication 7, d'une cellule selon la revendication 8 ou 9, d'un récepteur selon l'une quelconque des revendications 1 à 4, dans un test de criblage pour identifier des nouveaux médicaments.
  - Procédé pour identifier des ligands fonctionnels pour un récepteur selon l'une quelconque des revendications 1 à
     ledit procédé comprenant les étapes de
    - (a) introduction dans une cellule hôte appropriée:

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- (i) d'un ADN selon l'une quelconque des revendications 5 ou 6, et
- (ii) d'un gène indicateur approprié lié fonctionnellement à un élément de réponse pour la fonction hormonale (HRE), ledit HRE pouvant être activé par le domaine de liaison d'ADN du récepteur codé par ledit ADN;
- (b) mise en contact de ladite cellule hôte de (a) avec des ligands potentiels qui sont susceptibles de se lier au domaine de liaison du ligand du récepteur codé par ledit ADN de l'étape (a) ; et
- (c) suivi de l'expression du récepteur codé par ledit gène indicateur de l'étape (a).

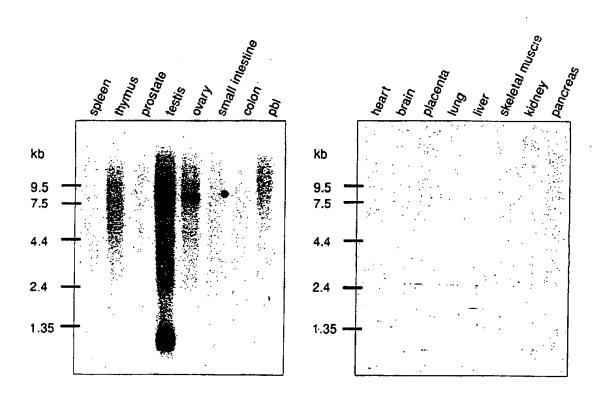
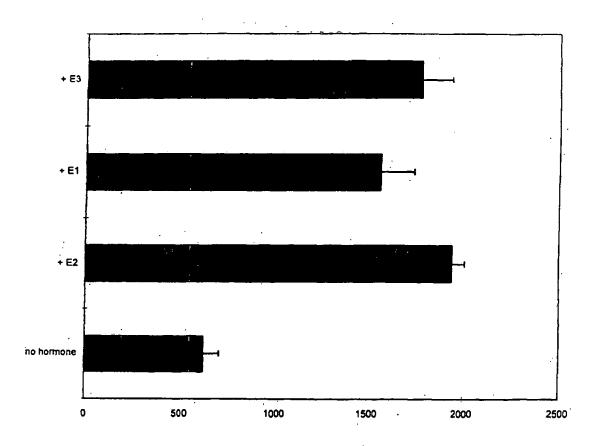


Figure 1



luciferase units

Fig. 2

# Transient transfection of CHO cells with Estrogen Receptors Alpha and Beta Incubation with Estradiol and ICI

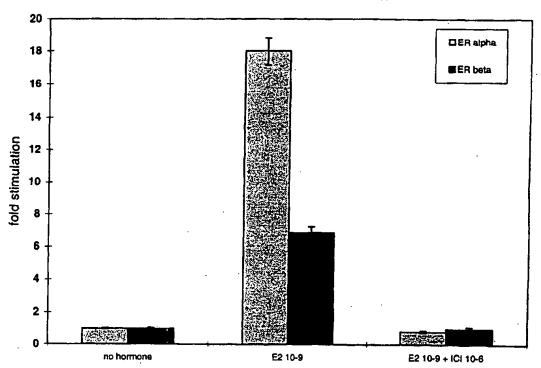


Figure 3

# ERα and ERβ RT PCR on tissue-representative cell lines

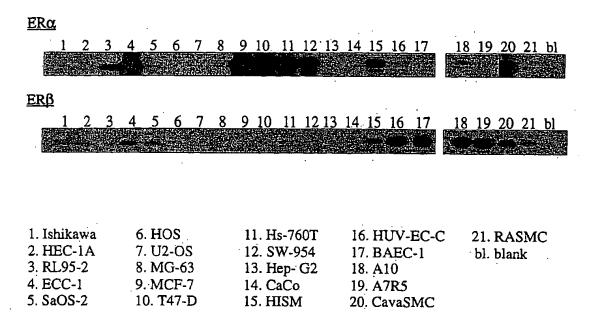
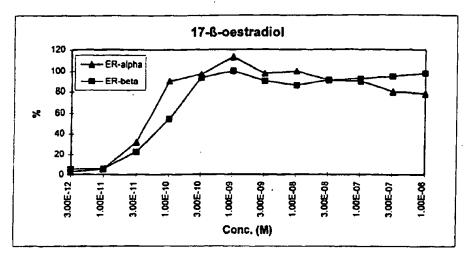
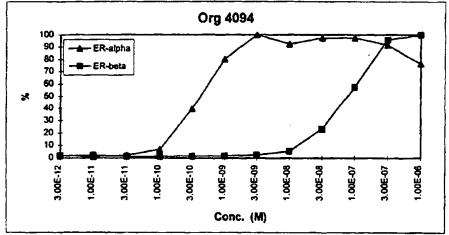


Figure 4





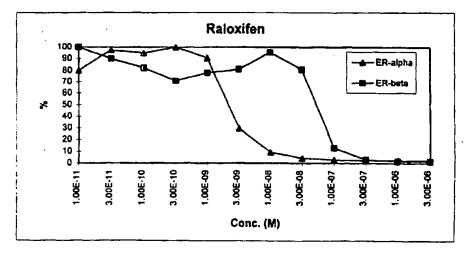


Figure 5